# Full resolution CTD profiles from R/V Hugh R. Sharp cruise HRS100808BW in 2010 (Marine Nitrogen Cycling by Stable Isotope Probing project)

Website: https://www.bco-dmo.org/dataset/3525

Version: August 29 2011 Version Date: 2011-08-29

### **Project**

» <u>Determining rates of group-specific phytoplankton and bacterial uptake of inorganic and organic nitrogen by</u> means of stable isotope techniques (Marine Nitrogen Cycling by Stable Isotope Probing)

## **Program**

» Ocean Carbon and Biogeochemistry (OCB)

Contributors	Affiliation	Role
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# **Dataset Description**

Full resolution profiles taken as part of study on uptake of phytoplankton and bacterial uptake of inorganic and organic nitrogen by means of stable isotope techniques.

## Methods & Sampling

Data were acquired using a Seabird SBE 9 CTD connected to an SBE 11 plus deck unit. The CTD was equipped with an SBE Oxygen sensor, a WET Labs ECO\_AFL fluorometer, a turbidity sensor, and a transmissometer. Seabird acquisition software Seasave V7.12 was used.

## **Data Processing Description**

Raw data were converted to ascii format using Seabird processing software. Derived properties include

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## **Data Files**

## File

CTD.csv(Comma Separated Values (.csv), 2.50 MB) MD5:6379913d140fae9e24ce1146ded17ba9

Primary data file for dataset ID 3525

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# **Parameters**

Parameter	Description	Units
cruise	Cruise identifier	dimensionless
date	Date of sample	YYYYMMDD
lon	longitude	decimal degrees
lat	latitude	decimal degrees
prmax	pressure max	decibars
cast	station identifier	dimensionless
temp	Temperature	degrees Celsius
temp2	Temperature from Secondary Sensor	degrees Celsius
sal	Salinity	dimensionless
sal2	Salinity from Secondary sensor	dimensionless
sigma_0	water potential density	kg m^-3
sigma_0_2	water potential density from secondary TC sensors	kg m^-3
O2_ml_L	dissolved oxygen concentration	ml per liter
O2_umol_kg	dissolved oxygen concentration	micromoles per kilogram
flvolt	fluorometer	volts
trans_v	transmissivity	volts
sound_vel	sound velocity	meters per second
beam_cp	beam attenuation	reciprocal meters
time_elapsed	Elapsed Time	seconds
press	Pressure	decibars

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# Instruments

Dataset- specific Instrument Name	CTD Sea-Bird 911
Generic Instrument Name	CTD Sea-Bird 911
	The Sea-Bird SBE 911 is a type of CTD instrument package. The SBE 911 includes the SBE 9 Underwater Unit and the SBE 11 Deck Unit (for real-time readout using conductive wire) for deployment from a vessel. The combination of the SBE 9 and SBE 11 is called a SBE 911. The SBE 9 uses Sea-Bird's standard modular temperature and conductivity sensors (SBE 3 and SBE 4). The SBE 9 CTD can be configured with auxiliary sensors to measure other parameters including dissolved oxygen, pH, turbidity, fluorescence, light (PAR), light transmission, etc.). More information from Sea-Bird Electronics.

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# **Deployments**

### HRS100808BW

Website	https://www.bco-dmo.org/deployment/58715
Platform	R/V Hugh R. Sharp
Start Date	2010-08-10
End Date	2010-08-16
Description	August 2010 Marine Nitrogen Cycling by Stable Isotope Probing cruise in Chespeake Bay, funded by: NSF OCE-0241310 Original cruise data are available from the NSF R2R data catalog

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## **Project Information**

Determining rates of group-specific phytoplankton and bacterial uptake of inorganic and organic nitrogen by means of stable isotope techniques (Marine Nitrogen Cycling by Stable Isotope Probing)

Coverage: Chesapeake Bay

From the NSF award abstract: The marine nitrogen (N) cycle involves a complex network of biological transformations among different inorganic and organic N reservoirs. Considerable progress has been made in defining N cycling processes in marine environments in recent years, but significant questions remain unanswered in part due to methodological limitations. Traditional tools for studying N cycling, for example, cannot accurately assess phytoplankton or bacteria specific N use in marine ecosystems. Therefore there is a need to develop new techniques and methodologies. The PIs of this project have recently made two important advances in this context: (1) a flowcytometric methodology (FCM) to separate phytoplankton from bacteria was applied to separately measure N uptake by these two groups. Prior methodologies relied on measurements of different size fractions, which always contain some degree of both phytoplankton and bacterial uptake. FCM allows for the distinct separation of bacterial versus phytoplankton N incorporation. (2) N-based DNA stable isotope probing (SIP) methodology has been adapted to interrogate N uptake in specific phytoplankton populations. DNA SIP can provide evidence for the uptake of an N source into a specific population of phytoplankton or bacteria. This methodology is in contrast to traditional measurements, which cannot make inferences about individual populations or species.

This project aims to apply these two methodological advances in order to obtain the next generation of N

uptake measurements. Phytoplankton and bacteria specific uptake rates will be measured via the FCM technique, and the individual groups or species of phytoplankton or bacteria will be interrogated for N uptake via DNA SIP. These tools will be applied across the well-characterized nutrient gradient found in Chesapeake Bay during one summer cruise and one winter cruise. Phytoplankton, bacterial, and archaeal populations will be characterized along the sampling transect via multiplexed pyrosequencing technology. N uptake will be measured for inorganic (NH4+, NO3-, and NO2-) and organic N sources (15N and 14C urea dual-labeled and amino acids) as substrates. The investigators hypothesize that phytoplankton will derive a larger percentage of their N nutrition from organic forms along the transect (i.e. North to South), as competition with bacteria for ammonium increases. DNA SIP will be applied to specific dominant phytoplankton and bacterial populations in order to investigate their N nutrition. By applying this unique combination of methodologies, the project will provide unprecedented community, group and species level resolution of N uptake in Chesapeake Bay and will furnish us with an improved understanding of N cycling in the Bay and marine systems as a whole.

Related Publication: Wawrik, B; Callaghan, AV; Bronk, DA. "Use of Inorganic and Organic Nitrogen by Synechococcus spp. and Diatoms on the West Florida Shelf as Measured Using Stable Isotope Probing," *APPLIED AND ENVIRONMENTAL MICROBIOLOGY*, v.75, 2009, p. 6662-6670. View record at Web of Science

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# **Program Information**

Ocean Carbon and Biogeochemistry (OCB)

Website: http://us-ocb.org/

Coverage: Global

The Ocean Carbon and Biogeochemistry (OCB) program focuses on the ocean's role as a component of the global Earth system, bringing together research in geochemistry, ocean physics, and ecology that inform on and advance our understanding of ocean biogeochemistry. The overall program goals are to promote, plan, and coordinate collaborative, multidisciplinary research opportunities within the U.S. research community and with international partners. Important OCB-related activities currently include: the Ocean Carbon and Climate Change (OCCC) and the North American Carbon Program (NACP); U.S. contributions to IMBER, SOLAS, CARBOOCEAN; and numerous U.S. single-investigator and medium-size research projects funded by U.S. federal agencies including NASA, NOAA, and NSF.

The scientific mission of OCB is to study the evolving role of the ocean in the global carbon cycle, in the face of environmental variability and change through studies of marine biogeochemical cycles and associated ecosystems.

The overarching OCB science themes include improved understanding and prediction of: 1) oceanic uptake and release of atmospheric CO2 and other greenhouse gases and 2) environmental sensitivities of biogeochemical cycles, marine ecosystems, and interactions between the two.

The OCB Research Priorities (updated January 2012) include: ocean acidification; terrestrial/coastal carbon fluxes and exchanges; climate sensitivities of and change in ecosystem structure and associated impacts on biogeochemical cycles; mesopelagic ecological and biogeochemical interactions; benthic-pelagic feedbacks on biogeochemical cycles; ocean carbon uptake and storage; and expanding low-oxygen conditions in the coastal and open oceans.

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## **Funding**

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-0960806
NSF Division of Ocean Sciences (NSF OCE)	OCE-0961900

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