

# Macroalgal contribution to NH<sub>4</sub><sup>+</sup> removal in tidepools from Cape Flattery, WA from 2010-2011 (Regenerated Nitrogen project)

**Website:** <https://www.bco-dmo.org/dataset/489420>

**Version:** 2014-11-04

## Project

» [The Role of Regenerated Nitrogen for Rocky Shore Productivity](#) (Regenerated Nitrogen)

Contributors	Affiliation	Role
<a href="#">Pather, Santhiska</a>	University of Massachusetts Dartmouth (UMASSD-SMAST)	Principal Investigator, Student
<a href="#">Altabet, Mark A.</a>	University of Massachusetts Dartmouth (UMASSD-SMAST)	Co-Principal Investigator
<a href="#">Pfister, Catherine</a>	University of Chicago	Co-Principal Investigator
<a href="#">Post, David</a>	Yale University	Co-Principal Investigator
<a href="#">Copley, Nancy</a>	Woods Hole Oceanographic Institution (WHOI BCO-DMO)	BCO-DMO Data Manager

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## Dataset Description

We evaluated the role of mussels by adding 15N-labeled NH<sub>4</sub><sup>+</sup> to an assemblage of tidepools where they were either present at natural abundance levels, or absent through manual removal. The role of phototrophs was separately examined by conducting these experiments both during the day and at night. The tidal height of pools varied between 1.2 to 1.5 m above Mean Lower Low Water (MLLW). Tidepools were thus isolated from each other as well as the nearshore environment during the low tide period when experiments were conducted. Each experiment included 4 to 5 mussel removal (MR) tidepools (since 2002) and 4 to 5 mussel control (MC) tidepools with natural mussel densities. In June 2010, we performed daytime NH<sub>4</sub><sup>+</sup> tracer experiments and in August 2010 nighttime experiments using the same tidepools. The following year (July 2011) these experiments were repeated with the addition of bottle incubations (see below) to evaluate the effects of suspended tidepool components and extended sampling for 6 days after the initial 15N addition to test for long-term retention of NH<sub>4</sub><sup>+</sup>. During the 2011 experiments, unforeseen rain reduced the salinity in some pools by up to 51%, and we have attempted to correct for the expected dilution of NH<sub>4</sub><sup>+</sup> in our tidepool rate calculations.

Because isotope enrichment levels were relatively low, we used the conventional delta notation instead of atom% to describe variations in 15N enrichment (where  $\delta\text{-}^{15}\text{N}\text{NH}_4^+ = \{(\text{15N:14N sample} \div \text{15N:14N standard}) - 1\} \times 1000\text{‰}$ , where the standard is atmospheric N<sub>2</sub>). Tracer labeled ammonium chloride (15NH<sub>4</sub>Cl) was added to the pools to approximate a 1000‰ enrichment in 2010 (doubling the 15N-NH<sub>4</sub><sup>+</sup> concentration) and a 2000‰ enrichment in 2011(tripling the 15N-NH<sub>4</sub><sup>+</sup> concentration). 15N natural abundance is only 0.365‰ and these tracer additions thus had a negligible effect on the overall NH<sub>4</sub><sup>+</sup> concentrations increasing them by only ~0.4‰ and ~0.8‰, respectively. Tidepool volumes were estimated spectrophotometrically using varying concentrations of food dye (Pfister 1995). Together with estimates of NH<sub>4</sub><sup>+</sup> concentration (from 2009 data), we estimated the tracer addition required to achieve the targeted 15N enrichments. However, the actual initial enrichments varied substantially, 684.4 - 2406.4‰ in 2010 and 781.4 - 3880.2‰ in 2011, likely due to error in tidepool volume estimation and natural variations in initial NH<sub>4</sub><sup>+</sup> concentrations. Fortunately, we sampled immediately following each tracer addition allowing for the determination of the true initial 15N

enrichment.

Prior to tracer addition at ebb tide, 100 mL of tidepool water was syringe-filtered (Whatman GF/F) into separate HDPE bottles for natural abundance  $^{15}\text{NH}_4^+$  and concentration determination. To each pool, tracer  $^{15}\text{NH}_4^+$  was then added and distributed by stirring with a stick. Water samples were immediately collected for measuring initial  $^{15}\text{N}$  enrichment and subsequently at 2, 4, and 6 hour intervals to determine isotope and concentration time courses. All water samples were frozen until analysis. Tidepool oxygen, pH, and temperature (Hach HQ4D) were also collected at ~ 2 h intervals throughout the experiment.

In 2011 we also assessed the contribution of the suspended microbial community to  $\text{NH}_4^+$  cycling by enclosing tidepool water in a 250 mL transparent polycarbonate incubation bottle. Following tracer addition, the bottle was filled, then left to float in the tidepool for the duration of the experiment. Samples from bottles were filtered as described both immediately after containment and at the end of the experiment (~6 h later).

We assessed macroalgal contribution to  $\text{NH}_4^+$  removal by transplanting two tidepool-dwelling algae species. *Prionitis sternbergii* were sampled 2 weeks prior to the experiment for baseline natural abundance  $^{15}\text{N}$  values and transplanted into the pools with Z-Spar Epoxy (Pfister 2007). On the day of the experiment, the red-alga, *Corallina vancouveriensis* from a single source patch, was also sampled for  $^{15}\text{N}$  natural abundance, inserted into pieces of Styrofoam, and floated in each pool.

At the end of each experiment (~6 h sampling point), we sampled tidepool particulate organic material (POM) by filtering through combusted GF/F filters until they clogged (~ 600 mL), comparing these samples with POM similarly sampled from the immediate nearshore. Floating *Corallina* spp. samples were collected into clean Eppendorf tubes, and similar sized pieces of *Prionitis* spp. were collected from each pool into clean foil packets.

We evaluated the extent of longer-term  $^{15}\text{N}$  tracer retention in 2011 by sampling tidepool water, POM and transplanted *Prionitis* 1, 3, and 6 days following tracer addition. We sampled at ebb tide and at again at slack water just prior to high tide on the first day after tracer addition (that is, 24 h later) and at slack water prior to high tide on Day 3 and 6.

## Relevant References:

2014. Pather, S., C. A. Pfister, M. Altabet, D. M. Post. Ammonium cycling in the rocky intertidal: remineralization, removal and retention. *Limnology and Oceanography* 59:361-372. [http://aslo.org/lo/toc/vol\\_59/issue\\_2/0361.htm](http://aslo.org/lo/toc/vol_59/issue_2/0361.htm)

DOI for this dataset: The role of regenerated nitrogen for rocky shore productivity, Cape Flattery, Washington, 2010 & 2011. Handle: <http://hdl.handle.net/1912/6420>. DOI:10.1575/1912/6420

## Related Datasets:

[filter tracer content](#)

[natural abundance N and C filter content](#)

[tidepool ammonium and mussels](#)

[tidepool incubation ammonium](#)

## Methods & Sampling

### Laboratory analysis:

$\text{NH}_4^+$  and  $\text{NO}_3^-$  concentrations were measured at the University of Washington Marine Chemistry Lab (methods from UNESCO 1994). Concentration values from 2011 were corrected for rain dilution using the change in salinity measured over the incubation period assuming no addition of  $\text{NH}_4^+$  or  $\text{NO}_3^-$  to the pools from the rainfall.

$\text{NH}_4^+$  isotopic composition was measured according to a modified version of Zhang et al. (2007) after isotope dilution to less than 500‰ to prevent isotopic contamination of the natural abundance-level mass spectrometer system. Briefly,  $\text{NH}_4^+$  is oxidized to nitrite using hypobromite then reduced to  $\text{N}_2\text{O}$  using acetic acid buffered sodium azide before analysis on an isotope ratio mass spectrometer (IRMS). In modification of the prior method, pre-existing  $\text{NO}_2^-$  was removed prior to hypobromite addition by reaction with sulfamic acid. To a 20 mL sample volume, 340  $\mu\text{L}$  20 mmol L<sup>-1</sup> sulfamic acid and 10  $\mu\text{L}$  10% HCl was added and allowed to react for 12 hours at room temperature. A second improvement was the addition of 6 mol L<sup>-1</sup> HCl to reduce the pH of the sample below 7 prior to the addition of an azide-100% acetic acid reagent. Isotope

determinations were made at U. Massachusetts Dartmouth using a GV IsoPrime IRMS, a custom purge-trap sample preparation system, and a CTC PAL autosampler. Reproducibility was better than  $\pm 0.5\%$ .

Filters and algal samples were dried at 60°C for 48 h and elemental and isotopic analyses were made at the University of Chicago and at Yale University. Samples were run using a Costech 4010 Elemental Analyzer combustion system coupled to a Thermo DeltaV Plus IRMS via a Thermo Conflo IV interface (University of Chicago), or using the same Elemental Analyzer coupled to a Thermo DeltaXP Advantage IRMS via a Thermo Conflo III interface (Yale University). Reproducibility was better than  $\pm 0.1\%$ .

## Data Processing Description

Raw  $\text{NH}_4^+$  isotope data were corrected for a background 'blank' using a simple mass balance equation (Zhang et al. 2007) to remove the contribution made to the signal height from reagent and diluent low nutrient seawater (LNSW)  $\text{NH}_4^+$  contamination. Data were then calibrated using a standard curve derived from co-analysis of international standards National Institute of Standards and Technology United States Geological Survey 25 ( $-29.4\%$ ) and 26 ( $+52.9\%$ ), and International Atomic Energy Agency N1 ( $0.5\%$ ) per Zhang et al. (2007). A final correction was made for the initial isotopic dilution.

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## Data Files

File
<b>seaweed_NH4.csv</b> (Comma Separated Values (.csv), 5.30 KB) MD5:c4f62d02805d24df0ad29c5654e6b492
Primary data file for dataset ID 489420

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## Parameters

Parameter	Description	Units
lat	latitude; north is positive	decimal degrees
lon	longitude; east is positive	decimal degrees
species	seaweed species in tidepool	unitless
station	tidepool identification number	unitless
treatment	mussels removed or present in tidepool	unitless
date	date samples collected in yyyyymmdd format	unitless
sample	sample identification number	unitless
d15N	nitrogen isotopic composition (delta 15N:N14) of the (mostly organic) material trapped on the filter	parts per thousand vs. VSMOW (Vienna Standard Mean Ocean Water)
N_pcent	percent nitrogen of filter	percent
C_pcent	percent carbon of filter	percent
site	sampling location	unitless

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## Instruments

<b>Dataset-specific Instrument Name</b>	CHN_EA
<b>Generic Instrument Name</b>	CHN Elemental Analyzer
<b>Dataset-specific Description</b>	Samples were run using a Costech 4010 Elemental Analyzer combustion system coupled to a Thermo DeltaV Plus IRMS via a Thermo Conflo IV interface (University of Chicago), or using the same Elemental Analyzer coupled to a Thermo DeltaXP Advantage IRMS via a Thermo Conflo III interface (Yale University). Reproducibility was better than $\pm 0.1\%$ . These instruments were used to look at the mass composition and isotopic signatures of the algal and filter material.
<b>Generic Instrument Description</b>	A CHN Elemental Analyzer is used for the determination of carbon, hydrogen, and nitrogen content in organic and other types of materials, including solids, liquids, volatile, and viscous samples.

<b>Dataset-specific Instrument Name</b>	IR Mass Spec
<b>Generic Instrument Name</b>	Isotope-ratio Mass Spectrometer
<b>Dataset-specific Description</b>	GV IsoPrime IRMS: Isotope determinations were made at U. Massachusetts Dartmouth using a GV IsoPrime IRMS, a custom purge-trap sample preparation system, and a CTC PAL autosampler. Reproducibility was better than $\pm 0.5\%$ .
<b>Generic Instrument Description</b>	The Isotope-ratio Mass Spectrometer is a particular type of mass spectrometer used to measure the relative abundance of isotopes in a given sample (e.g. VG Prism II Isotope Ratio Mass-Spectrometer).

<b>Dataset-specific Instrument Name</b>	Nutrient Autoanalyzer
<b>Generic Instrument Name</b>	Nutrient Autoanalyzer
<b>Dataset-specific Description</b>	The nutrient autoanalyzer at UWashington was used to determine the nutrient concentrations in the water. Analyses and calibration follow the protocols of the WOCE Hydrographic Program using a Technicon AAll system. For more information, see <a href="http://www.ocean.washington.edu/story/Marine+Chemistry+Laboratory">http://www.ocean.washington.edu/story/Marine+Chemistry+Laboratory</a>
<b>Generic Instrument Description</b>	Nutrient Autoanalyzer is a generic term used when specific type, make and model were not specified. In general, a Nutrient Autoanalyzer is an automated flow-thru system for doing nutrient analysis (nitrate, ammonium, orthophosphate, and silicate) on seawater samples.

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## Deployments

### Cape\_Flaterry\_2011

<b>Website</b>	<a href="https://www.bco-dmo.org/deployment/488878">https://www.bco-dmo.org/deployment/488878</a>
<b>Platform</b>	Cape_Flaterry
<b>Start Date</b>	2010-06-26
<b>End Date</b>	2011-07-22
<b>Description</b>	Nitrogen studies in tidepools.

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## Project Information

### The Role of Regenerated Nitrogen for Rocky Shore Productivity (Regenerated Nitrogen)

**Website:** <http://pfisterlab.uchicago.edu>

**Coverage:** coastal northeast Pacific Ocean

*NSF Award Abstract:*

A fundamental and persistent question in a multitude of ecosystems is the extent to which new versus regenerated nutrients support ecosystem productivity. In coastal marine systems, nitrate derived from upwelling (= new nitrogen) and ammonium regeneration in coastal waters and sediments (= regenerated nitrogen) are major nitrogen sources that fuel coastal ocean productivity. Because inorganic nitrogen availability clearly regulates production in a large number of areas, understanding nitrogen supply is essential. In open coast regions away from river mouths, nitrate inputs are determined by large-scale physical processes promoting upwelling of deep, nutrient-rich water including wind direction and intensity. In contrast, regenerated nitrogen (mainly ammonium) is generally the result of local animal and microbial processes. Along marine rocky shores, where upwelling is typically used as a proxy for productivity, we know very little about the dynamics of regenerated nutrients and their potential contribution to productivity at larger scales; only upwelling is typically used as a proxy for productivity. Associations of the abundant California mussel, *Mytilus californianus*, with water nutrients, algal productivity, stable isotope signatures, and microbial genetics indicate potentially strong regeneration of nitrogen by these animals and suggest an important secondary role of nitrifying microbes affiliated with these animals.

In this project, the investigators will quantify the relative contribution of regenerated nitrogen on rocky shores through censuses and experiments across a gradient of mussel abundance. They will use stable nitrogen and oxygen isotopes of ammonium, nitrite, and nitrate to disentangle the contribution of different biological processes versus upwelling to the nitrogen supply and uptake of rocky shore regions. This includes both natural abundance and tracer addition studies.

**Broader Impacts.** Regenerated nitrogen supply, as opposed to new nitrogen via upwelling, is a local process dependent upon an intact animal community. However, mussels and other nearshore animals may be particularly vulnerable to a changing thermal environment, toxic algal blooms, and ocean acidification. Given the dramatic changes to the coastal nitrogen cycle in recent years, and potential changes to currents, upwelling, ocean chemistry, and El Niño frequencies portended by global changes to our climate, we to know the relative effect of local versus larger scale oceanic events on the nitrogen cycle. The proposed work links biological interactions in situ with its implications for coastal productivity.

In addition to expected publications in high quality journals, educational activities will continue to focus on graduate and undergraduate education and mentoring. The proposal will fund two graduate students and two undergraduates per year. The PI's will work closely with government (Olympic Marine National Sanctuary) and tribal (Makah Tribe) representatives to communicate this research. We will also work with Makah Museum Board of Trustees and the Makah Higher Education Committee to identify Makah students as research assistants. All three PI's teach broadly across their respective campuses, instructing almost every type of undergraduate major.

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## Funding

Funding Source	Award
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-0928232</a>
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-0928015</a>
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-0928152</a>

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