

Dissolved nutrients from phyto and microzooplankton experiments from the RVIB Nathaniel B. Palmer NBP0601 cruise in the Ross Sea, Southern Ocean from 2005-2006 (CORSACS project, Antarctic microzooplankton project)

Website: <https://www.bco-dmo.org/dataset/504863>

Version:

Version Date: 2014-02-20

Project

» [Controls of Ross Sea Algal Community Structure](#) (CORSACS)

» [Rising climatic temperatures impact on antarctic microzooplankton growth and grazing](#) (Antarctic microzooplankton)

Program

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Dataset Description

Experimental Design

Experiments were conducted during the CORSACS (Controls On Ross Sea Algal Community Structure) expedition in January 2006 to the Ross Sea, Antarctica, onboard the RVIB Nathaniel B. Palmer (cruise NBP-0601). Water was collected at 75.00S, 177.36E using a trace metal clean towed-intake surface water Teflon diaphragm pumping system (Bruland et al., 2005). Water was prescreened through acid-washed 200µm Nitex mesh to eliminate large zooplankton and collected into a 50-L mixing carboy. Collected water was gently mixed and dispensed into 12 4.5-L and 12 2.7-L acid washed trace metal clean clear polycarbonate bottles for incubation. Half of the bottles were spiked with 1.0nM FeCl₃ (final concentration) at the beginning of the experiment. Bottles were incubated in two temperature controlled deck-board incubators (Feng et al., 2009; Hare et al., 2007). Incubators were screened to 18‰ of $\rho_{\sigma-t}$ using two layers of neutral density filter. One incubator was kept at ambient temperature (0 deg C), while the temperature in the other was gradually increased to 4 deg C over the course of 24 h. Bottles were incubated for seven days. The 4.5-L bottles were sampled daily and the 2.7-L bottles were only sampled on the final day of the experiment. All sampling occurred under a laminar flow hood using trace metal clean techniques.

References

Bruland, K.W., E.L. Rue, G.J. Smith, and G.R. DiTullio. 2005. Iron, macronutrients and diatom blooms in the Peru upwelling regime: brown and blue waters of Peru. *Marine Chemistry* 93: 81-103.

Feng, Y., C.E. Hare, K. Leblanc, G.R. DiTullio, P.A. Lee, S.W. Wilhelm, J. Sun, J.M. Rose, N. Nemcek, I. Benner, and D.A. Hutchins. 2009. The effects of increased pCO₂ and temperature on the North Atlantic Spring Bloom: I. The phytoplankton community and biogeochemical response. *Marine Ecology Progress Series* 388: 13-25.

Hare, C.E., K. Leblanc, G.R. DiTullio, R.M. Kudela, Y. Zhang, P.A. Lee, S.F. Riseman, and D.A. Hutchins. 2007. Consequences of increased temperature and CO₂ for phytoplankton community structure in the Bering Sea. *Marine Ecology Progress Series* 352: 9-16.

Methods & Sampling

Dissolved nutrient Sampling:

Samples for dissolved nutrients were 0.2 um filtered and immediately analyzed using a Lachat QuikChem 8000 Flow Injection Analysis System. The precision of the nutrient analyses were as follows: phosphate 0.7%, nitrite 2.5%, nitrate + nitrite 1.0%, silicic acid 0.5%.

References:

Bruland, K.W., E.L. Rue, G.J. Smith, and G.R. DiTullio. 2005. Iron, macronutrients and diatom blooms in the Peru upwelling regime: brown and blue waters of Peru. *Marine Chemistry* 93: 81-103.

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Buck, K.N., M.C. Lohan, C.J.M. Berger, and K.W. Bruland. 2007. Dissolved iron speciation in two distinct river plumes and an estuary: implications for riverine iron supply. *Limnology and Oceanography* 52: 843-855.

Donat, J.R., and K.W. Bruland. 1988. Direct determination of dissolved cobalt and nickel in seawater by differential pulse cathodic stripping voltammetry preceded by adsorptive collection of cyclohexane-1,2,-dione dioxime complexes. *Analytical Chemistry* 60.

Ellwood, M., and C.M.G. Van den Berg. 2000. Zinc speciation in the Northeastern Atlantic Ocean. *Marine Chemistry* 68: 295-306.

Solorzano, L., and J.H. Sharp. 1980. Determination of total dissolved phosphorus and particulate phosphorus in natural waters. *Limnology and Oceanography* 25: 756-760.

Data Processing Description

BCO-DMO Processing Notes:

- File was sorted by treatment
- Added lat,lon values of original water sampling location to file
- Added BCO-DMO header lines
- Parameter names were edited to conform with BCO-DMO convention
- Changed value at bottle 5,Day 2,Treatment LTHF from 'lost' to 'nd'
- Changed value at bottles 10, 11, 12 from Day 7,Treatment HTHF from '

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Data Files

File
nuts_disslv_ant1.csv (Comma Separated Values (.csv), 4.75 KB) MD5:0543de38efe1db991eb24042194316c4
Primary data file for dataset ID 504863

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Parameters

Parameter	Description	Units
lat	Latitude component of geographic position where water was sampled.	decimal degrees
lon	Longitude component of geographic position where water was sampled.	decimal degrees
treatment	Experimental conditions varied during the experiment. Four treatments were used: High temperature, high iron (HTHF); High temperature, low iron (HTLF); Low temperature, high iron (LTHF); Low temperature, low iron (LTLF)	dimensionless
day	Sampling day during experiment. The experiment was conducted during January, 2006.	dimensionless
bottle	Experimental bottle number.	dimensionless
PO4	Dissolved Phosphate.	micromolar
NO2	Dissolved Nitrite.	micromolar
NO2_NO3	Nitrate plus Nitrite.	micromolar
NH4	Ammonium.	micromolar
SiO2	Silicon dioxide.	micromolar

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Instruments

Dataset-specific Instrument Name	FIA
Generic Instrument Name	Flow Injection Analyzer
Dataset-specific Description	Samples for dissolved nutrients were analyzed using a Lachat QuikChem 8000 Flow Injection Analysis System.
Generic Instrument Description	An instrument that performs flow injection analysis. Flow injection analysis (FIA) is an approach to chemical analysis that is accomplished by injecting a plug of sample into a flowing carrier stream. FIA is an automated method in which a sample is injected into a continuous flow of a carrier solution that mixes with other continuously flowing solutions before reaching a detector. Precision is dramatically increased when FIA is used instead of manual injections and as a result very specific FIA systems have been developed for a wide array of analytical techniques.

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Deployments

NBP0601

Website	https://www.bco-dmo.org/deployment/57985
Platform	RVIB Nathaniel B. Palmer
Report	http://data.bco-dmo.org/CORSACS/cruises/Dunbar_Hydrography_report_NBP0601.pdf
Start Date	2005-12-17
End Date	2006-01-30
Description	This was the first of two Controls of Ross Sea Algal Community Structure (CORSACS) project cruises and was funded by the NSF Office of Polar Programs. The NBP0601 cruise was conducted in the Ross Sea in December 2005 and January 2006, Ross Sea, ca. 65.21°S-78.65°S, 164.98°E-164.70°W, and supported by NSF research grant, OPP-0338097. The 'Science Plan and Project Description' document includes details of the cruise sampling strategy. Related Files: Science Plan and Project Descriptions (PDF file)Cruise track map (PDF file)Photo of Ice Breaker Nathaniel B. Palmer on station near Beaufort Island (JPG image) Related Sites: MGDS catalog: http://www.marine-geo.org/tools/search/entry.php?id=NBP0601

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Project Information

Controls of Ross Sea Algal Community Structure (CORSACS)

Website: <http://www.whoi.edu/sites/corsacs>

Coverage: Ross Sea Southern Ocean

Project summary

The Controls of Ross Sea Algal Community Structure (CORSACS) project was funded by the NSF Office of Polar Programs as "Collaborative Research: Interactive Effects of Iron, Light and Carbon Dioxide on Phytoplankton Community Dynamics in the Ross Sea". Two cruises were completed in 2006 to investigate the interactions between the primary productivity of the Ross Sea and pCO₂, iron and other trace elements. Data sets of carbon, nutrient, metal, and biological measurements will be reported.

The main objective in the proposed research was to investigate the relative importance and potential interactive effects of iron, light and CO₂ levels in structuring algal assemblages and growth rates in the Ross Sea. The investigators hypothesized that the interaction of these three variables largely determines the bottom-up control on these two dominant Southern Ocean phytoplankton taxa. While grazing and other loss processes are important variables in determining the relative dominance of these two taxa, the CORSACS research project was designed to focus on the bottom-up control mechanisms. It is important to understand such environmentally-driven taxonomic shifts in primary production, since they are expected to impact the fixation and export of carbon and nutrients, and the production of DMS, thus potentially providing both positive and negative feedbacks on climate.

The CORSACS investigators considered a range of ambient iron, light and pCO₂ levels that span those typically observed in the Ross Sea during the growing season. That is, dissolved iron ranging from ~0.1 nM (low iron) to greater than 1 nM (high iron) (Fitzwater et al. 2000; Sedwick et al. 2000); mean irradiance (resulting from vertical mixing/self shading) ranging from less than 10% I₀ (low light) to greater than 40% (high light) (Arrigo et al., 1998, 1999), possibly adjusted based on field observations during the CORSACS cruises; and pCO₂ ranging (Sweeney et al. 2001) from ~150 ppm (low CO₂) to the probable higher levels of pCO₂ - 750 ppm as a conservative estimate - that are likely to be attained later this century due to anthropogenic perturbation of the global carbon cycle (IPCC, 2001).

From the information previously available from both field observations and experiments, the investigators

formulated the following specific hypotheses regarding the interactive role of iron, light and CO₂ in regulating algal composition in the Ross Sea: diatoms bloom in the southern Ross Sea only under optimum conditions of high iron, light and pCO₂; colonial *Phaeocystis* dominate under conditions of high iron with either (or both) low light or low pCO₂; and solitary *Phaeocystis* are predominant under conditions of low iron with either (or both) low light or low pCO₂.

References:

Fitzwater, S.E., K.S. Johnson, R.M. Gordon, K.H. Coale, and W.O. Smith, Jr. (2000). Trace metal concentrations in the Ross Sea and their relationship with nutrients and growth. *Deep-Sea Research II*, 47: 3159-3179.

Martin JH, Gordon RM, Fitzwater SE. Iron in Antarctic waters. *Nature* 1990 ;345(6271):156-158. Martin JH. 1990. Glacial-interglacial CO₂ change: The iron hypothesis. *Paleoceanography* 5(1):1-13

P. N. Sedwick, G. R. DiTullio, and D. J. Mackey, Iron and manganese in the Ross Sea, Antarctica: Seasonal iron limitation in Antarctic shelf waters, *Journal of Geophysical Research*, 105 (C5), 11,321-11,336, 2000.

Sweeney, C. K. Arrigo, and G. van Gijken (2001). Prediction of seasonal changes in surface pCO₂ in the Ross Sea, Antarctica using ocean color satellite data. 2001 Annual AGU meeting, San Fransisco, CA Dec. 10-15.

IPCC, 2001: *Climate Change 2001: Synthesis Report*. A Contribution of Working Groups I, II, and III to the Third Assessment Report of the Intergovernmental Panel on Climate Change [Watson, R.T. and the Core Writing Team (eds.)]. Cambridge University Press, Cambridge, United Kingdom, and New York, NY, USA, 398 pp.

Publications

Saito, M. A., Goepfert, T. J., Noble, A. E., Bertrand, E. M., Sedwick, P. N., and DiTullio, G. R.: A seasonal study of dissolved cobalt in the Ross Sea, Antarctica: micronutrient behavior, absence of scavenging, and relationships with Zn, Cd, and P, *Biogeosciences*, 7, 4059-4082, doi:10.5194/bg-7-4059-2010, 2010 (<http://www.biogeosciences.net/7/4059/2010/bg-7-4059-2010.html>)

Bertrand EM, Saito MA, Lee PA, Dunbar RB, Sedwick PN and DiTullio GR (2011) Iron limitation of a springtime bacterial and phytoplankton community in the Ross Sea: implications for vitamin B12 nutrition. *Front. Microbio.* 2:160. doi: 10.3389/fmicb.2011.00160 (http://www.frontiersin.org/Aquatic_Microbiology/10.3389/fmicb.2011.00160/abstract)

Rising climatic temperatures impact on antarctic microzooplankton growth and grazing (Antarctic microzooplankton)

Coverage: Ross Sea

The investigator will examine to what extent rising climatic temperatures impact antarctic microzooplankton growth and grazing, and to what extent such an impact would modulate top-down control of phytoplankton growth in cold waters. The experimental part of the proposed work would take place in the Ross Sea, a permanently cold ecosystem, and the location of annual large-scale blooms of both diatoms and *Phaeocystis* antarctica. Changing climate regimes may alter current microzooplankton grazing rates on these blooms either directly through temperature increases or indirectly through algal community shifts. Complementary laboratory experiments on cultures of Antarctic microzooplankton will be conducted to determine the individual and combined effects of temperature and carbon dioxide levels on growth and grazing.

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Program Information

Ocean Carbon and Biogeochemistry (OCB)

Website: <http://us-ocb.org/>

Coverage: Global

The Ocean Carbon and Biogeochemistry (OCB) program focuses on the ocean's role as a component of the global Earth system, bringing together research in geochemistry, ocean physics, and ecology that inform on and advance our understanding of ocean biogeochemistry. The overall program goals are to promote, plan, and coordinate collaborative, multidisciplinary research opportunities within the U.S. research community and with international partners. Important OCB-related activities currently include: the Ocean Carbon and Climate Change (OCCC) and the North American Carbon Program (NACP); U.S. contributions to IMBER, SOLAS, CARBOOCEAN; and numerous U.S. single-investigator and medium-size research projects funded by U.S. federal agencies including NASA, NOAA, and NSF.

The scientific mission of OCB is to study the evolving role of the ocean in the global carbon cycle, in the face of environmental variability and change through studies of marine biogeochemical cycles and associated ecosystems.

The overarching OCB science themes include improved understanding and prediction of: 1) oceanic uptake and release of atmospheric CO₂ and other greenhouse gases and 2) environmental sensitivities of biogeochemical cycles, marine ecosystems, and interactions between the two.

The OCB Research Priorities (updated January 2012) include: ocean acidification; terrestrial/coastal carbon fluxes and exchanges; climate sensitivities of and change in ecosystem structure and associated impacts on biogeochemical cycles; mesopelagic ecological and biogeochemical interactions; benthic-pelagic feedbacks on biogeochemical cycles; ocean carbon uptake and storage; and expanding low-oxygen conditions in the coastal and open oceans.

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Funding

Funding Source	Award
NSF Antarctic Sciences (NSF ANT)	PLR-0528715

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