Eukaryote transcriptome assemblies and associated metadata available on iMicrobe (P-ExpEv project)

Website: https://www.bco-dmo.org/dataset/646128 Data Type: experimental Version: Version Date: 2016-05-20

Project

» Impacts of Evolution on the Response of Phytoplankton Populations to Rising CO2 (P-ExpEv)

Program

» <u>Science, Engineering and Education for Sustainability NSF-Wide Investment (SEES): Ocean Acidification</u> (formerly CRI-OA) (SEES-OA)

Contributors	Affiliation	Role
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Coverage

Spatial Extent: N:78.5922 E:166.75 S:-64.7667 W:-179.23

Dataset Description

This dataset includes links to eukaryote transcriptome assemblies and associated metadata for species grown under replete nutrients and either modern or future CO2 concentrations. Further information on the overarching project and experiment details (e.g., nutrient concentrations, growth medium, GenBank BioSample IDs, etc) are available on iMicrobe MMETSP site: <u>http://data.imicrobe.us/project/view/104</u> or by clicking on the accession numbers listed below.

Accession Species/condition

MMETSP0053
MMETSP0057Prorocentrum minimum (Pmi): replete; modern CO2MMETSP0057
MMETSP0088Prorocentrum minimum (Pmi): replete; future CO2MMETSP0092
MMETSP0093Chaetoceros affinis (Caf): replete; modern CO2MMETSP0093
MMETSP0097Alexandrium monilatum (Amo): replete; modern CO2MMETSP0097
MMETSP0143Alexandrium monilatum (Amo): replete; future CO2MMETSP0143
MMETSP0147Chyrsochromulina polylepis (Cpo): replete; modern CO2

MMETSP0292 Heterosigma akashiwo (Hak): replete;modern CO2 MMETSP0296 Heterosigma akashiwo (Hak): replete; future CO2 MMETSP1363 Gephyrocapsa oceanica (Goc): replete;modern CO2 MMETSP1364 Gephyrocapsa oceanica (Goc): replete; future CO2

Methods & Sampling

The workflow for transcriptome assembly is described on the MMETSP website and is available at: <u>http://marinemicroeukaryotes.org/home/faq</u>. Sequencing was done by the National Center for Genome Resources (NCGR).

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Data Files

File	
MMETSP_data.csv(Comma Separated Values (.csv), 1.72 KB) MD5:5e08b3166bd653228d79ef4be6275ea9	
Primary data file for dataset ID 646128	

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Parameters

Parameter	Description	Units
external_link	link to iMicrobe data repository	text
brief_desc	brief description of link	text

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Instruments

Dataset- specific Instrument Name	
Generic Instrument Name	Automated DNA Sequencer
Dataset- specific Description	Pacific Biosciences RS and Illumina HiSeq 2000 at NCGR
Generic Instrument Description	General term for a laboratory instrument used for deciphering the order of bases in a strand of DNA. Sanger sequencers detect fluorescence from different dyes that are used to identify the A, C, G, and T extension reactions. Contemporary or Pyrosequencer methods are based on detecting the activity of DNA polymerase (a DNA synthesizing enzyme) with another chemoluminescent enzyme. Essentially, the method allows sequencing of a single strand of DNA by synthesizing the complementary strand along it, one base pair at a time, and detecting which base was actually added at each step.

Deployments

Dynrman_2013		
Website	https://www.bco-dmo.org/deployment/692263	
Platform	LDEO	
Start Date	2013-01-01	
End Date	2017-03-31	
Description	Phytoplankton studies	

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Project Information

Impacts of Evolution on the Response of Phytoplankton Populations to Rising CO2 (P-ExpEv)

Coverage: Experiment housed in laboratories at Michigan State University

Note: This project is also affiliated with the <u>NSF BEACON Center for the Study of Evolution in Action</u>.

Project Description from NSF Award:

Human activities are driving up atmospheric carbon dioxide concentrations at an unprecedented rate, perturbing the ocean's carbonate buffering system, lowering oceanic pH, and changing the concentration and composition of dissolved inorganic carbon. Recent studies have shown that this ocean acidification has many short-term effects on phytoplankton, including changes in carbon fixation among others. These physiological changes could have profound effects on phytoplankton metabolism and community structure, with concomitant effects on Earth's carbon cycle and, hence, global climate. However, extrapolation of present understanding to the field are complicated by the possibility that natural populations might evolve in response to their changing environments, leading to different outcomes than those predicted from short-term studies. Indeed, evolution experiments demonstrate that microbes are often able to rapidly adapt to changes in the environment, and that beneficial mutations are capable of sweeping large populations on time scales relevant to predictions of environmental dynamics in the coming decades. This project addresses two major areas of uncertainty for phytoplankton populations with the following questions:

1) What adaptive mutations to elevated CO2 are easily accessible to extant species, how often do they arise, and how large are their effects on fitness?

2) How will physical and ecological interactions affect the expansion of those mutations into standing populations?

This study will address these questions by coupling experimental evolution with computational modeling of ocean biogeochemical cycles. First, cultured unicellular phytoplankton, representative of major functional groups (e.g. cyanobacteria, diatoms, coccolithophores), will be evolved under simulated year 2100 CO2 concentrations. From these experiments, estimates will be made of a) the rate of beneficial mutations, b) the magnitude of fitness gains conferred by these mutations, and c) secondary phenotypes (i.e., trade-offs) associated with these mutations, assayed using both physiological and genetic approaches. Second, an existing numerical model of the global ocean system will be modified to a) simulate the effects of changing atmospheric CO2 concentrations on ocean chemistry, and b) allow the introduction of CO2-specific adaptive mutants into the extant populations of virtual phytoplankton. The model will be used to explore the ecological and biogeochemical impacts of beneficial mutations in realistic environmental situations (e.g. resource availability, predation, etc.). Initially, the model will be applied to idealized sensitivity studies; then, as experimental results become available, the implications of the specific beneficial mutations observed in our experiments will be explored.

This interdisciplinary study will provide novel, transformative understanding of the extent to which evolutionary processes influence phytoplankton diversity, physiological ecology, and carbon cycling in the near-future ocean. One of many important outcomes will be the development and testing of nearly-neutral genetic markers useful for competition studies in major phytoplankton functional groups, which has applications well beyond

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Program Information

Science, Engineering and Education for Sustainability NSF-Wide Investment (SEES): Ocean Acidification (formerly CRI-OA) (SEES-OA)

Website: <u>https://www.nsf.gov/funding/pgm_summ.jsp?pims_id=503477</u>

Coverage: global

NSF Climate Research Investment (CRI) activities that were initiated in 2010 are now included under Science, Engineering and Education for Sustainability NSF-Wide Investment (SEES). SEES is a portfolio of activities that highlights NSF's unique role in helping society address the challenge(s) of achieving sustainability. Detailed information about the SEES program is available from NSF (<u>https://www.nsf.gov/funding/pgm_summ.jsp?</u> <u>pims_id=504707</u>).

In recognition of the need for basic research concerning the nature, extent and impact of ocean acidification on oceanic environments in the past, present and future, the goal of the SEES: OA program is to understand (a) the chemistry and physical chemistry of ocean acidification; (b) how ocean acidification interacts with processes at the organismal level; and (c) how the earth system history informs our understanding of the effects of ocean acidification on the present day and future ocean.

Solicitations issued under this program:

<u>NSF 10-530</u>, FY 2010-FY2011 <u>NSF 12-500</u>, FY 2012 <u>NSF 12-600</u>, FY 2013 <u>NSF 13-586</u>, FY 2014 NSF 13-586 was the final solicitation that will be released for this program.

PI Meetings:

<u>1st U.S. Ocean Acidification PI Meeting</u>(March 22-24, 2011, Woods Hole, MA) <u>2nd U.S. Ocean Acidification PI Meeting</u>(Sept. 18-20, 2013, Washington, DC) 3rd U.S. Ocean Acidification PI Meeting (June 9-11, 2015, Woods Hole, MA – Tentative)

NSF media releases for the Ocean Acidification Program:

Press Release 10-186 NSF Awards Grants to Study Effects of Ocean Acidification

Discovery Blue Mussels "Hang On" Along Rocky Shores: For How Long?

<u>Discovery nsf.gov - National Science Foundation (NSF) Discoveries - Trouble in Paradise: Ocean Acidification</u> <u>This Way Comes - US National Science Foundation (NSF)</u>

<u>Press Release 12-179 nsf.gov - National Science Foundation (NSF) News - Ocean Acidification: Finding New</u> <u>Answers Through National Science Foundation Research Grants - US National Science Foundation (NSF)</u>

Press Release 13-102 World Oceans Month Brings Mixed News for Oysters

<u>Press Release 13-108 nsf.gov - National Science Foundation (NSF) News - Natural Underwater Springs Show</u> <u>How Coral Reefs Respond to Ocean Acidification - US National Science Foundation (NSF)</u>

<u>Press Release 13-148 Ocean acidification: Making new discoveries through National Science Foundation</u> <u>research grants</u>

<u>Press Release 13-148 - Video nsf.gov - News - Video - NSF Ocean Sciences Division Director David Conover</u> <u>answers questions about ocean acidification. - US National Science Foundation (NSF)</u> <u>Press Release 14-010 nsf.gov - National Science Foundation (NSF) News - Palau's coral reefs surprisingly</u> resistant to ocean acidification - US National Science Foundation (NSF)

<u>Press Release 14-116 nsf.gov - National Science Foundation (NSF) News - Ocean Acidification: NSF awards</u> <u>\$11.4 million in new grants to study effects on marine ecosystems - US National Science Foundation (NSF)</u>

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Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-1314336

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