

Grazing experiment 4a: Short term data for low-high pCO₂ acclimated *Rhodomonas* sp. cultures ingested by *Gyrodinium* grazers (E Hux Response to pCO₂ project)

Website: <https://www.bco-dmo.org/dataset/668661>

Data Type: experimental

Version:

Version Date: 2016-12-06

Project

» [Planktonic interactions in a changing ocean: Biological responses of *Emiliana huxleyi* to elevated pCO₂ and their effects on microzooplankton](#) (E Hux Response to pCO₂)

Contributors	Affiliation	Role
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Dataset Description

Related Reference:

Still, Kelly Ann, Microzooplankton grazing, growth and gross growth efficiency are affected by pCO₂ induced changes in phytoplankton biology. (Masters Thesis) Western Washington University. <http://cedar.wvu.edu/cgi/viewcontent.cgi?article=1490&context=wwuet>

Methods & Sampling

The phytoplankton *Rhodomonas* sp. CCMP 755 was grown semi-continuously in atmosphere controlled chambers at three different CO₂ treatment concentrations; Ambient (400ppmv), Moderate (750ppmv), and High (1000ppmv). Cultures were diluted daily starting day 4 with pre-equilibrated media containing f/50 nutrients. On day 9 *Rhodomonas* cells from the treatments were fed to starved *Gyrodinium dominans* cells for 15, 45, and 90 minutes. A 24-hour time point was also taken but scored at either “1” for containing any food or “0” for no food in the vacuole. During each sampling time point, 20 ml of experiment volume was removed, fixed with 0.5% glutaraldehyde and stained with DAPI. Samples were stored at 4° C for 12 hours to allow time for the DAPI to stain. This volume was filtered onto a 5 µm pore size polycarbonate filter with, which was then put on a slide with immersion oil and kept frozen until evaluation. Slides were evaluated under 400x using an epi-fluorescent microscope under blue-light excitation. The first 100 microzooplankton on each slide were assessed for each replicate/treatment, and individual prey cells were counted in the grazer food vacuole.

Data Processing Description

These data are unprocessed counts of the Rhodomonas cells ingested by each Gyrodinium grazer.

BCO-DMO Processing Notes:

- added conventional header with dataset name, PI name, version date
- modified parameter names to conform with BCO-DMO naming conventions
- nd (no data) was entered into all blank cells

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Data Files

File
expt4a_Gyro_grazing.csv (Comma Separated Values (.csv), 9.49 KB) MD5:0cc4139b82e904305db22a2409efe523
Primary data file for dataset ID 668661

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Parameters

Parameter	Description	Units
grazer_analyzed	sample identifier: individual grazer analyzed	unitless
amb_15min_A	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 15 minute grazing period; replicate A	cells
amb_15min_B	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 15 minute grazing period; replicate B	cells
amb_15min_C	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 15 minute grazing period; replicate C	cells
amb_45min_A	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 45 minute grazing period; replicate A	cells
amb_45min_B	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 45 minute grazing period; replicate B	cells
amb_45min_C	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 45 minute grazing period; replicate C	cells
amb_90min_A	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 90 minute grazing period; replicate A	cells

amb_90min_B	Rhodomonas cells counted inside the grazer: ambient pCO ₂ level; 90 minute grazing period; replicate B	cells
amb_90min_C	Rhodomonas cells counted inside the grazer: ambient pCO ₂ level; 90 minute grazing period; replicate C	cells
amb_24hr_A	Rhodomonas cells counted inside the grazer: ambient pCO ₂ level; 24 hour grazing period; replicate A	cells
amb_24hr_B	Rhodomonas cells counted inside the grazer: ambient pCO ₂ level; 24 hour grazing period; replicate B	cells
amb_24hr_C	Rhodomonas cells counted inside the grazer: ambient pCO ₂ level; 24 hour grazing period; replicate C	cells
mod_15min_A	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 15 minute grazing period; replicate A	cells
mod_15min_B	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 15 minute grazing period; replicate B	cells
mod_15min_C	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 15 minute grazing period; replicate C	cells
mod_45min_A	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 45 minute grazing period; replicate A	cells
mod_45min_B	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 45 minute grazing period; replicate B	cells
mod_45min_C	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 45 minute grazing period; replicate C	cells
mod_90min_A	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 90 minute grazing period; replicate A	cells
mod_90min_B	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 90 minute grazing period; replicate B	cells
mod_90min_C	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 90 minute grazing period; replicate C	cells
mod_24hr_A	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 24 hour grazing period; replicate A	cells

mod_24hr_B	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 24 hour grazing period; replicate B	cells
mod_24hr_C	Rhodomonas cells counted inside the grazer: moderate pCO ₂ level; 24 hour grazing period; replicate C	cells
high_15min_A	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 15 minute grazing period; replicate A	cells
high_15min_B	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 15 minute grazing period; replicate B	cells
high_15min_C	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 15 minute grazing period; replicate C	cells
high_45min_A	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 45 minute grazing period; replicate A	cells
high_45min_B	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 45 minute grazing period; replicate B	cells
high_45min_C	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 45 minute grazing period; replicate C	cells
high_90min_A	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 90 minute grazing period; replicate A	cells
high_90min_B	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 90 minute grazing period; replicate B	cells
high_90min_C	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 90 minute grazing period; replicate C	cells
high_24hr_A	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 24 hour grazing period; replicate A	cells
high_24hr_B	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 24 hour grazing period; replicate B	cells
high_24hr_C	Rhodomonas cells counted inside the grazer: high pCO ₂ level; 24 hour grazing period; replicate C	cells

Instruments

Dataset-specific Instrument Name	epi-fluorescent microscope under blue-light excitation
Generic Instrument Name	Fluorescence Microscope
Dataset-specific Description	For cell counts
Generic Instrument Description	Instruments that generate enlarged images of samples using the phenomena of fluorescence and phosphorescence instead of, or in addition to, reflection and absorption of visible light. Includes conventional and inverted instruments.

Deployments

Lab_Olson_B

Website	https://www.bco-dmo.org/deployment/521277
Platform	WWU
Start Date	2011-03-31
End Date	2016-09-15
Description	laboratory experiments

Project Information

Planktonic interactions in a changing ocean: Biological responses of *Emiliana huxleyi* to elevated pCO₂ and their effects on microzooplankton (E Hux Response to pCO₂)

Description from NSF award abstract:

The calcifying Haptophyte *Emiliana huxleyi* appears to be acutely sensitive to the rising concentration of ocean pCO₂. Documented responses by *E. huxleyi* to elevated pCO₂ include modifications to their calcification rate and cell size, malformation of coccoliths, elevated growth rates, increased organic carbon production, lowering of PIC:POC ratios, and elevated production of the active climate gas DMS. Changes in these parameters are mechanisms known to elicit alterations in grazing behavior by microzooplankton, the oceans dominant grazer functional group. The investigators hypothesize that modifications to the physiology and biochemistry of calcifying and non-calcifying Haptophyte *Emiliana huxleyi* in response to elevated pCO₂ will precipitate alterations in microzooplankton grazing dynamics. To test this hypothesis, they will conduct controlled laboratory experiments where several strains of *E. huxleyi* are grown at several CO₂ concentrations. After careful characterization of the biochemical and physiological responses of the *E. huxleyi* strains to elevated pCO₂, they will provide these strains as food to several ecologically-important microzooplankton and document grazing dynamics. *E. huxleyi* is an ideal organism for the study of phytoplankton and microzooplankton responses to rising anthropogenic CO₂, the effects of which in the marine environment are called ocean

acidification; *E. huxleyi* is biogeochemically important, is well studied, numerous strains are in culture that exhibit variation in the parameters described above, and they are readily fed upon by ecologically important microzooplankton.

The implications of changes in microzooplankton grazing for carbon cycling, specifically CaCO₃ export, DMS production, nutrient regeneration in surface waters, and carbon transfer between trophic levels are profound, as this grazing, to a large degree, regulates all these processes. *E. huxleyi* is a model prey organism because it is one of the most biogeochemically influential global phytoplankton. It forms massive seasonal blooms, contributes significantly to marine inorganic and organic carbon cycles, is a large producer of the climatically active gas DMS, and is a source of organic matter for trophic levels both above and below itself. The planned controlled study will increase our knowledge of the mechanisms that drive patterns of change between trophic levels, thus providing a wider array of tools necessary to understand the complex nature of ocean acidification field studies, where competing variables can confound precise interpretation.

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Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-0961229

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