

# Short term data for low-high pCO<sub>2</sub> acclimated *Rhodomonas* sp. cultures ingested by *Favella* grazers (E Hux Response to pCO<sub>2</sub> project)

**Website:** <https://www.bco-dmo.org/dataset/668747>

**Data Type:** experimental

**Version:**

**Version Date:** 2016-12-06

## Project

» [Planktonic interactions in a changing ocean: Biological responses of \*Emiliana huxleyi\* to elevated pCO<sub>2</sub> and their effects on microzooplankton](#) (E Hux Response to pCO<sub>2</sub>)

Contributors	Affiliation	Role
<a href="#">Olson, M Brady</a>	Western Washington University (WWU)	Principal Investigator
<a href="#">Love, Brooke</a>	Western Washington University (WWU)	Co-Principal Investigator
<a href="#">Strom, Suzanne</a>	Western Washington University (WWU)	Co-Principal Investigator
<a href="#">Still, Kelly Ann</a>	Western Washington University (WWU)	Student
<a href="#">Copley, Nancy</a>	Woods Hole Oceanographic Institution (WHOI BCO-DMO)	BCO-DMO Data Manager

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## Dataset Description

### Related Reference:

Still, Kelly Ann, Microzooplankton grazing, growth and gross growth efficiency are affected by pCO<sub>2</sub> induced changes in phytoplankton biology. (Masters Thesis) Western Washington University. <http://cedar.wvu.edu/cgi/viewcontent.cgi?article=1490&context=wwuet>

## Methods & Sampling

The phytoplankton *Rhodomonas* sp. CCMP 755 was grown semi-continuously in atmosphere controlled chambers at three different CO<sub>2</sub> treatment concentrations; Ambient (400ppmv), Moderate (750ppmv), and High (1000ppmv). Cultures were diluted daily starting day 4 with pre-equilibrated media containing f/50 nutrients. On day 9 *Rhodomonas* cells from the treatments were fed to starved *Favella ehrenbergii* cells for 15, 30, and 45 minutes. A 24-hour time point was also taken but scored at either "1" for containing any food or "0" for no food in the vacuole. During each sampling time point, 20 ml of experiment volume was removed, fixed with 0.5% glutaraldehyde and stained with DAPI. Samples were stored at 4° C for 12 hours to allow time for the DAPI to stain. This volume was filtered onto a 20 µm pore size polycarbonate filter with, which was then put on a slide with immersion oil and kept frozen until evaluation. Slides were evaluated under 400x using an epi-fluorescent microscope under blue-light excitation. The first 100 microzooplankton on each slide were assessed for each replicate/treatment, and individual prey cells were counted in the grazer food vacuole.

## Data Processing Description

These data are unprocessed counts of the Rhodomonas cells ingested by each Favella grazer.

### BCO-DMO Processing Notes:

- added conventional header with dataset name, PI name, version date
- modified parameter names to conform with BCO-DMO naming conventions
- nd (no data) was entered into all blank cells

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## Data Files

File
<b>expt4b_Fav_grazing.csv</b> (Comma Separated Values (.csv), 10.41 KB) MD5:9f62a69c57a98a1a8fea51b16d09c822
Primary data file for dataset ID 668747

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## Parameters

Parameter	Description	Units
Grazer_Analyzed	sample identifier: individual grazer analyzed	unitless
amb_15min_A	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 15 minute grazing period; replicate A	cells
amb_15min_B	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 15 minute grazing period; replicate B	cells
amb_15min_C	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 15 minute grazing period; replicate C	cells
amb_45min_A	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 45 minute grazing period; replicate A	cells
amb_45min_B	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 45 minute grazing period; replicate B	cells
amb_45min_C	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 15 minute grazing period; replicate C	cells
amb_90min_A	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 90 minute grazing period; replicate A	cells
amb_90min_B	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 90 minute grazing period; replicate B	cells
amb_90min_C	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 90 minute grazing period; replicate C	cells
amb_24hr_A	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 24 hour grazing period; replicate A	cells
amb_24hr_B	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 24 hour grazing period; replicate B	cells
amb_24hr_C	Rhodomonas cells counted inside the grazer: ambient pCO2 level; 24 hour grazing period; replicate C	cells

mod_15min_A	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 15 minute grazing period; replicate A	cells
mod_15min_B	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 15 minute grazing period; replicate B	cells
mod_15min_C	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 15 minute grazing period; replicate C	cells
mod_45min_A	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 45 minute grazing period; replicate A	cells
mod_45min_B	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 45 minute grazing period; replicate B	cells
mod_45min_C	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 15 minute grazing period; replicate C	cells
mod_90min_A	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 90 minute grazing period; replicate A	cells
mod_90min_B	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 90 minute grazing period; replicate B	cells
mod_90min_C	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 90 minute grazing period; replicate C	cells
mod_24hr_A	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 24 hour grazing period; replicate A	cells
mod_24hr_B	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 24 hour grazing period; replicate B	cells
mod_24hr_C	Rhodomonas cells counted inside the grazer: moderate pCO2 level; 24 hour grazing period; replicate C	cells
high_15min_A	Rhodomonas cells counted inside the grazer: high pCO2 level; 15 minute grazing period; replicate A	cells
high_15min_B	Rhodomonas cells counted inside the grazer: high pCO2 level; 15 minute grazing period; replicate B	cells
high_15min_C	Rhodomonas cells counted inside the grazer: high pCO2 level; 15 minute grazing period; replicate C	cells
high_45min_A	Rhodomonas cells counted inside the grazer: high pCO2 level; 45 minute grazing period; replicate A	cells
high_45min_B	Rhodomonas cells counted inside the grazer: high pCO2 level; 45 minute grazing period; replicate B	cells
high_45min_C	Rhodomonas cells counted inside the grazer: high pCO2 level; 15 minute grazing period; replicate C	cells
high_90min_A	Rhodomonas cells counted inside the grazer: high pCO2 level; 90 minute grazing period; replicate A	cells
high_90min_B	Rhodomonas cells counted inside the grazer: high pCO2 level; 90 minute grazing period; replicate B	cells
high_90min_C	Rhodomonas cells counted inside the grazer: high pCO2 level; 90 minute grazing period; replicate C	cells
high_24hr_A	Rhodomonas cells counted inside the grazer: high pCO2 level; 24 hour grazing period; replicate A	cells
high_24hr_B	Rhodomonas cells counted inside the grazer: high pCO2 level; 24 hour grazing period; replicate B	cells
high_24hr_C	Rhodomonas cells counted inside the grazer: high pCO2 level; 24 hour grazing period; replicate C	cells

## Instruments

<b>Dataset-specific Instrument Name</b>	epi-fluorescent microscope under blue-light excitation
<b>Generic Instrument Name</b>	Fluorescence Microscope
<b>Dataset-specific Description</b>	For cell counts
<b>Generic Instrument Description</b>	Instruments that generate enlarged images of samples using the phenomena of fluorescence and phosphorescence instead of, or in addition to, reflection and absorption of visible light. Includes conventional and inverted instruments.

## Deployments

### Lab\_Olson\_B

<b>Website</b>	<a href="https://www.bco-dmo.org/deployment/521277">https://www.bco-dmo.org/deployment/521277</a>
<b>Platform</b>	WWU
<b>Start Date</b>	2011-03-31
<b>End Date</b>	2016-09-15
<b>Description</b>	laboratory experiments

## Project Information

### **Planktonic interactions in a changing ocean: Biological responses of *Emiliana huxleyi* to elevated pCO<sub>2</sub> and their effects on microzooplankton (E Hux Response to pCO<sub>2</sub>)**

#### ***Description from NSF award abstract:***

The calcifying Haptophyte *Emiliana huxleyi* appears to be acutely sensitive to the rising concentration of ocean pCO<sub>2</sub>. Documented responses by *E. huxleyi* to elevated pCO<sub>2</sub> include modifications to their calcification rate and cell size, malformation of coccoliths, elevated growth rates, increased organic carbon production, lowering of PIC:POC ratios, and elevated production of the active climate gas DMS. Changes in these parameters are mechanisms known to elicit alterations in grazing behavior by microzooplankton, the oceans dominant grazer functional group. The investigators hypothesize that modifications to the physiology and biochemistry of calcifying and non-calcifying Haptophyte *Emiliana huxleyi* in response to elevated pCO<sub>2</sub> will precipitate alterations in microzooplankton grazing dynamics. To test this hypothesis, they will conduct controlled laboratory experiments where several strains of *E. huxleyi* are grown at several CO<sub>2</sub> concentrations. After careful characterization of the biochemical and physiological responses of the *E. huxleyi* strains to elevated pCO<sub>2</sub>, they will provide these strains as food to several ecologically-important microzooplankton and document grazing dynamics. *E. huxleyi* is an ideal organism for the study of phytoplankton and microzooplankton responses to rising anthropogenic CO<sub>2</sub>, the effects of which in the marine environment are called ocean acidification; *E. huxleyi* is biogeochemically important, is well studied, numerous strains are in culture that exhibit variation in the parameters described above, and they are readily fed upon by ecologically important

microzooplankton.

The implications of changes in microzooplankton grazing for carbon cycling, specifically CaCO<sub>3</sub> export, DMS production, nutrient regeneration in surface waters, and carbon transfer between trophic levels are profound, as this grazing, to a large degree, regulates all these processes. *E. huxleyi* is a model prey organism because it is one of the most biogeochemically influential global phytoplankton. It forms massive seasonal blooms, contributes significantly to marine inorganic and organic carbon cycles, is a large producer of the climatically active gas DMS, and is a source of organic matter for trophic levels both above and below itself. The planned controlled study will increase our knowledge of the mechanisms that drive patterns of change between trophic levels, thus providing a wider array of tools necessary to understand the complex nature of ocean acidification field studies, where competing variables can confound precise interpretation.

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## Funding

Funding Source	Award
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-0961229</a>

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