

# N isotopic composition of Phenylalanine and Glutamic Acid from a number of organisms, demonstrating new HPLC protocol for precise isotopic measurements

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## Project

» [The Use of Nitrogen Isotopes of Amino Acids To Understand Marine Sedimentary  \$^{15}\text{N}\$  Records](#) (Amino Acid Sediment  $^{15}\text{N}$ )

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## Abstract

N isotopic composition of Phenylalanine and Glutamic Acid from a number of organisms, demonstrating new HPLC protocol for precise isotopic measurements.

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## Dataset Description

N isotopic composition of Phenylalanine and Glutamic Acid from a number of organisms.

These data were published in:

Broek, T, and McCarthy, M.D. (2014) A New Approach to  $\delta^{15}\text{N}$  compound-specific amino acid trophic position measurements: preparative high pressure liquid chromatography technique for purifying underivatized amino acids for stable isotope analysis. *Limnology and Oceanography Methods*, 12, 2014, 840-852.  
doi:[10.4319/lom.2014.12.840](https://doi.org/10.4319/lom.2014.12.840)

## Methods & Sampling

### Methodology described in Broek & McCarthy (2014):

#### AA standards

Standard L-AA powders were purchased from Alfa Aesar and Acros Organics and used to prepare individual liquid standards (0.05 M), which were then combined as an equimolar mixture of 16 individual AAs ("16 AA Standard") for developing separations. The 16 AA Standard contained the proteinaceous AAs: glycine (Gly), L-alanine (Ala), L-arginine (Arg), L-aspartic acid (Asp), L-glutamic acid (Glu), L-histidine (His), L-isoleucine (Ile), L-leucine (Leu), L-lysine (Lys), D/L-methionine (Met), L-phenylalanine (Phe), L-proline (Pro), L-serine (Ser), L-threonine (Thr), L-valine (Val); and nonprotein AA nor-leucine (Nle), which is commonly used as an internal standard (Popp et al. 2007; McCarthy et al. 2013). The  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values for dry standards were determined by standard EA-IRMS at the University of California, Santa Cruz Stable Isotope Laboratory (UCSC-

SIL) following standard protocols (<http://es.ucsc.edu/~silab>). Average precision of EA-IRMS  $\delta^{15}\text{N}$  standard values was  $0.11 \pm 0.07$  ‰. Additionally, a commercially available equimolar AA standard mixture "Pierce Amino Acid Standard H" (Pierce H)(Thermo Scientific) containing the same AAs as the "16 AA Standard" with the exception of the nonprotein AA Nle and addition of the proteinaceous AAs cysteine (Cys) and tyrosine (Tyr) was used to construct individual calibration curves, so as to verify relative molar abundance of individual AAs in natural samples.

### Sample preparation:

The cyanobacteria sample (*Spirulina Sp.*) was obtained as a bulk commercial dry powder (Spirulina Pacifica, Nutrex Hawaii, Kailua-Kona, HI). This same sample has been used previously as a McCarthy laboratory internal quality control standard, and its CSI-AA values have been measured repeatedly by GC-C-IRMS, allowing an investigation of the long-term accuracy and precision of the GC-C-IRMS instrument.

Coastal mussel (*Mytilus Californianus*) sample was collected in 2012 from Santa Cruz, CA. The mussel was previously dissected, and the adductor muscle tissue removed and lyophilized prior to storage. We used a subsample of adductor muscle collected for a prior study (Vokhshoori and McCarthy 2014) hydrolyzing the bulk lyophilized adductor muscle tissue directly without lipid extraction.

The deep-sea bamboo coral (genus *Isidella*) sample was previously collected in 2007 from Monterey Bay, CA, USA (36 44.6538N, 122 2.2329W, 870.2 m) (Hill, *pers. comm.* 2011). A proteinaceous node was separated from the calcium carbonate skeleton and oven dried (60 degrees C, 24 h).

White sea bass muscle tissue was subsampled from an incidental recreational catch in 2007, landed from Santa Cruz Island, Channel Islands, CA (J. Patterson, *pers. comm.* 2007). Fish muscle tissue was also lyophilized prior to hydrolysis.

Harbor seal blood was collected in May-June 2007 from a wild animal in Tomales Bay, CA (38 13.9N, 122 58.1W) under NMFS Research Permit no. 555-1565. Blood serum was purified, lipid extracted, and lyophilized prior to hydrolysis, as described previously (Germain et al. 2011).

For all sample types, proteinaceous material was hydrolyzed by adding 40-50 mg of bulk dry sample to an 8 mL glass vial, followed by 5 mL of 6 N hydrochloric acid (HCl) at room temperature. The vials were flushed with nitrogen gas, sealed, and allowed to hydrolyze under standard conditions (110 degrees C, 20 h). Hydrolysis under acidic conditions quantitatively deaminates asparagine (Asn) to aspartic acid, and glutamine (Gln) to glutamic acid (Barrett 1985). Therefore, in this protocol (and all others based on acid hydrolysis), measured Glu in fact represents Gln+Glu, and measured Asp represents Asp+Asn. We note that while the abbreviations Glx and Asx are sometimes used to denote these combined Gln+Glu and Asp+Asn fractions, we have elected to simply use Asp and Glu as abbreviations, as defined above, in order to correspond better with prior TP<sub>CSIA</sub> literature. Additionally, acid hydrolysis is known to destroy cysteine (Cys), precluding it from analysis (Barrett 1985). Resulting hydrolysates were dried to completion under nitrogen gas and brought up in 0.1 N HCl to a final concentration of 1 mg tissue/ 100 L HCl. Approximately 75% of each of the resulting mixtures was reserved for HPLC/EA-IRMS analysis, and the remaining material was dried to completion for derivatization and subsequent GC-C-IRMS analysis.

### GC-C-IRMS Analysis:

Trifluoroacetyl isopropyl ester (TFA-IP) AA derivatives were prepared using standardized lab protocols, as described previously (McCarthy et al. 2013). Briefly, hydrolyzed samples were esterified in 300  $\mu\text{L}$  1:5 mixture of acetyl chloride:2-propanol (110 degrees C, 60 minutes). The resulting amino acid isopropyl esters were then acylated in 350  $\mu\text{L}$  1:3 mixture of dichloromethane (DCM):trifluoroacetic acid anhydride (100 degrees C, 15 minutes). Derivatized AAs were dissolved in DCM to a final ratio of 1 mg of original proteinaceous material to 50  $\mu\text{L}$  DCM. Isotopic analysis was conducted on a Thermo Trace GC Ultra (Thermo Fisher Scientific, West Palm Beach, FL, USA) coupled via a Thermo GC IsoLink to a ThermoFinnigan DeltaPlus XP isotope ratio monitoring mass spectrometer (Thermo Fisher Scientific). Derivatives (1 L) were injected (injector temp. 250 degrees C constant) onto an Agilent DB-5 column (50 m x 0.32 mm ID x 0.52  $\mu\text{m}$  film thickness, Agilent Technologies, Inc., Santa Clara, CA, USA), with a He carrier flow rate of 2 mL/min (constant-flow). Separations were achieved with a four-ramp oven program: 52 deg C, 2 min hold; ramp 1 = 15 deg C /min to 75 deg C, hold for 2 min; ramp 2 = 4 deg C /min to 185 deg C, hold for 2 min; ramp 3 = 4 deg C /min to 200 deg C; ramp 4 = 30 deg C /min to 240 deg C, hold for 5 min. This method allows for the determination of 11-15 AAs depending on derivatization efficiency and instrument sensitivity. Values are typically obtained for Gly, Ala, Glu, Ile, Leu, Phe, Pro, Ser, Thr, Val, Nle, and Lys. Values for Met, His and Arg are obtained only in some samples, depending on concentration and derivatization efficiency. For  $\delta^{15}\text{N}$  AA values, samples were analyzed in quadruplicate ( $n=4$ ) with bracketed lab AA isotopic standard mix for subsequent standard offset and drift corrections. Corrections based on authentic external standards were applied using previously published protocols (McCarthy et al.

2013).

### **HPLC/EA-IRMS:**

Liquid chromatographic separations were conducted using a Shimadzu HPLC system (Shimadzu Scientific Instruments, Inc., Columbia, MD, USA) equipped with: system controller (SCL-10A vp), degasser (DGU-20A5), 2 pumps (LC-20AD), autosampler (SIL-20A) with an adjustable injection volume of 0.1-100  $\mu$ L, and coupled to a Shimadzu automated fraction collector (FRC-20A). An adjustable flow splitter (Analytical Sales and Services, Inc., Pompton Plains, NJ, USA) was used inline following the chromatography column to direct ~15% of the flow to a SEDERE (Alfortville, France) evaporative light scattering detector (ELSD-LT II, Sedex 85LT) for peak detection and quantitation. A semi-preparative scale SiELC Primesep A column (10 x 250 mm, 100 angstrom pore size, 5  $\mu$ m particle size; SiELC Technologies Ltd., Prospect Heights, IL, USA) was used for amino acid purification. The Primesep A column used here is a reverse-phase semi-preparative scale column embedded with strong acidic ion-pairing groups. Such mixed phase columns have been developed specifically for the separation of charged organic compounds as the acidic sites in the stationary phase interact with the charged functional groups and provide additional retention mechanisms to increase chromatographic separation potential. For a more detailed description of the retention mechanisms of the Primesep A column see (McCullagh et al. 2006; 2010).

Typically, 75-100  $\mu$ L of sample solution was loaded onto the HPLC instrument. A binary solvent ramp program was used consisting of 0.1% trifluoroacetic acid (TFA) in HPLC grade water (aqueous phase) and 0.1% TFA in acetonitrile (organic phase). The final solvent ramp program used for optimal separation was as follows: starting with 100% aqueous / 0% organic; increased from 0 to 0.5% organic from 0-30 minutes; increased to 15% organic from 30-35 minutes; increased to 22.5% from 35-70 minutes; increased to 30% from 70-95 minutes; held at 30% until 140 minutes. The column was then cleaned and equilibrated by increasing to 100% and holding for 20 minutes; then decreasing to 50% and holding for an additional 15 minutes; then decreasing to 0% and holding until the method ends at 180 minutes. A flow rate ramp is also employed in which the total flow rate is held at 2.5 mL/minute for 0-30 minutes; increased to 4.5 mL/minute from 30-35 minutes; held at 4.5 mL/min from 35-170 minutes; then decreased back to 2.5 mL/minute from 170-175 minutes and held until the completion of the analysis.

Purified AAs were collected into 3.5 mL tubes via the automated fraction collector using time-based collections, and then transferred to 20 mL glass vials. The solvent was removed under vacuum using a Jouan centrifugal evaporator (Societe Jouan, Saint-Herblain, France) at a chamber temperature of 60 degrees C. Dry AA residues were then re-dissolved into a small volume (~30 L) of 0.1 N HCl, transferred into pre-ashed tin (Sn) EA capsules, and dried to completion in a 60 degrees C oven for 12 hours. Capsules were then pressed into cubes and analyzed for  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values by EA-IRMS. EA-IRMS analysis was conducted in the UCSC shared Stable Isotope Laboratory facility (UCSC-SIL), using an EA-IRMS analyzer dedicated to smaller samples. This system uses a Carlo Erba CHNS-O EA1108-Elemental Analyzer, interfaced via a Thermo Finnigan Gasbench II device to a Thermo Finnigan Delta Plus XP isotope ratio mass spectrometer (Thermo Fisher Scientific), configured after Polissar et al. (2009). For AAs in this study, we found that  $\leq 100$  nmol quantities of purified AA material could be routinely measured using this instrument, although as discussed below a standard EA configuration could also equally be used. Raw EA-IRMS  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values were corrected for instrument drift and size effects using AA isotopic standards and standard correction protocols used by the UCSC-SIL (<http://es.ucsc.edu/~silab>).

### **Data Processing Description**

BCO-DMO Processing:

- modified parameter names to conform with BCO-DMO naming conventions;
- replaced missing data/blanks with "nd" ("no data")

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### **Data Files**

File
<b>mol_percent.csv</b> (Comma Separated Values (.csv), 1.12 KB) MD5:8a2988eb54fe1a0579cdd25f565dd23c
Primary data file for dataset ID 712090

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## Related Publications

Broek, T. A. B., & McCarthy, M. D. (2014). A new approach to  $\delta^{15}\text{N}$  compound-specific amino acid trophic position measurements: preparative high pressure liquid chromatography technique for purifying underivatized amino acids for stable isotope analysis. *Limnology and Oceanography: Methods*, 12(12), 840–852.

doi:[10.4319/lom.2014.12.840](https://doi.org/10.4319/lom.2014.12.840)

*Results*

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## Parameters

Parameter	Description	Units
sample_type	Description of the sample type.	unitless
amino_acid	Amino acid	unitless
mol_pcnt_hplc	Relative amino acid abundance; determined by HPLC-ELSD. Average precision: +/- 1	percent (%)
mol_pcnt_hplc_stdev	Standard deviation of relative amino acid abundance determined by HPLC-ELSD.	percent (%)
mol_pcnt_GCC	Relative amino acid abundance; determined by GC-C-IRMS. Average precision: +/- 1	percent (%)
mol_pcnt_GCC_stdev	Standard deviation of relative amino acid abundance determined by GC-C-IRMS.	percent (%)

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## Instruments

<b>Dataset-specific Instrument Name</b>	Carlo Erba CHNS-O EA1108-elemental analyzer
<b>Generic Instrument Name</b>	Elemental Analyzer
<b>Dataset-specific Description</b>	EA-IRMS analysis was conducted in the UCSC shared Stable Isotope Laboratory facility (UCSC-SIL), using an EA-IRMS analyzer dedicated to smaller samples. This system uses a Carlo Erba CHNS-O EA1108-Elemental Analyzer, interfaced via a Thermo Finnigan Gasbench II device to a Thermo Finnigan Delta Plus XP isotope ratio mass spectrometer (Thermo Fisher Scientific).
<b>Generic Instrument Description</b>	Instruments that quantify carbon, nitrogen and sometimes other elements by combusting the sample at very high temperature and assaying the resulting gaseous oxides. Usually used for samples including organic material.

<b>Dataset-specific Instrument Name</b>	Thermo Trace GC Ultra
<b>Generic Instrument Name</b>	Gas Chromatograph
<b>Dataset-specific Description</b>	Isotopic analysis was conducted on a Thermo Trace GC Ultra (Thermo Fisher Scientific, West Palm Beach, FL, USA) coupled via a Thermo GC IsoLink to a ThermoFinnigan DeltaPlus XP isotope ratio monitoring mass spectrometer (Thermo Fisher Scientific).
<b>Generic Instrument Description</b>	Instrument separating gases, volatile substances, or substances dissolved in a volatile solvent by transporting an inert gas through a column packed with a sorbent to a detector for assay. (from SeaDataNet, BODC)

<b>Dataset-specific Instrument Name</b>	Agilent DB-5 column
<b>Generic Instrument Name</b>	Gas Chromatograph
<b>Dataset-specific Description</b>	Derivatives (1 L) were injected (injector temp. 250 degrees C constant) onto an Agilent DB-5 column.
<b>Generic Instrument Description</b>	Instrument separating gases, volatile substances, or substances dissolved in a volatile solvent by transporting an inert gas through a column packed with a sorbent to a detector for assay. (from SeaDataNet, BODC)

<b>Dataset-specific Instrument Name</b>	Shimadzu HPLC
<b>Generic Instrument Name</b>	High-Performance Liquid Chromatograph
<b>Dataset-specific Description</b>	Liquid chromatographic separations were conducted using a Shimadzu HPLC system (Shimadzu Scientific Instruments, Inc., Columbia, MD, USA) equipped with: system controller (SCL-10A vp), degasser (DGU-20A5), 2 pumps (LC-20AD), autosampler (SIL-20A) with an adjustable injection volume of 0.1-100 $\mu$ L, and coupled to a Shimadzu automated fraction collector (FRC-20A).
<b>Generic Instrument Description</b>	A High-performance liquid chromatograph (HPLC) is a type of liquid chromatography used to separate compounds that are dissolved in solution. HPLC instruments consist of a reservoir of the mobile phase, a pump, an injector, a separation column, and a detector. Compounds are separated by high pressure pumping of the sample mixture onto a column packed with microspheres coated with the stationary phase. The different components in the mixture pass through the column at different rates due to differences in their partitioning behavior between the mobile liquid phase and the stationary phase.

<b>Dataset-specific Instrument Name</b>	ThermoFinnigan DeltaPlus XP isotope ratio monitoring mass spectrometer
<b>Generic Instrument Name</b>	Isotope-ratio Mass Spectrometer
<b>Dataset-specific Description</b>	Isotopic analysis was conducted on a Thermo Trace GC Ultra (Thermo Fisher Scientific, West Palm Beach, FL, USA) coupled via a Thermo GC IsoLink to a ThermoFinnigan DeltaPlus XP isotope ratio monitoring mass spectrometer (Thermo Fisher Scientific).
<b>Generic Instrument Description</b>	The Isotope-ratio Mass Spectrometer is a particular type of mass spectrometer used to measure the relative abundance of isotopes in a given sample (e.g. VG Prism II Isotope Ratio Mass-Spectrometer).

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## Project Information

### The Use of Nitrogen Isotopes of Amino Acids To Understand Marine Sedimentary $\delta^{15}\text{N}$ Records (Amino Acid Sediment $\delta^{15}\text{N}$ )

**Coverage:** California Margin , Santa Barbara Basin , CA current system, Eastern Tropical Pacific

The bioavailability of nutrients plays a crucial role in oceanic biological productivity, the carbon cycle, and climate change. The global ocean inventory of nitrogen (N) is determined by the balance of N-fixation (sources) and denitrification (sinks). In this three-year project, a researcher from the University of California, Santa Cruz, will focus on developing compound-specific N isotope ( $\delta^{15}\text{N}$ ) analysis of amino acids as a new tool for understanding N source and transformation of organic matter in paleo-reservoirs. The offsets in the isotopic ratios of individual amino acid groups may yield information about trophic transfer, heterotrophic microbial reworking, and autotrophic versus heterotrophic sources. By measuring and comparing the bulk and amino acid  $\delta^{15}\text{N}$  in size-fractionated samples from plankton tows, sediments traps, and multi-cores in oxic and suboxic depositional environments, the researcher will: (1) Provide a proxy of the  $\delta^{15}\text{N}$  of average exported photoautotrophic organic matter; and (2) Provide a new level of detail into sedimentary organic N degradation and preservation.

Broader impacts:

This project will improve understanding of the fundamental underpinnings and behaviors of  $\delta^{15}\text{N}$  amino acid patterns and how they behave in contrasting sedimentary environments, while also developing a potential paleoceanographic proxy. Funding will support a graduate student and undergraduate research at the institution. The researcher will also conduct community outreach in the form of a workshop/tutorial on the proxy development.

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## Funding

Funding Source	Award
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-1131816</a>

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