

# Growth rates and equivalent spherical diameters of *Heterosigma akashiwo* after temperature transition

**Website:** <https://www.bco-dmo.org/dataset/783500>

**Data Type:** experimental

**Version:** 2

**Version Date:** 2020-01-06

## Project

» [Quantifying Temperature Dependence In Growth & Grazing Rates of Planktonic Herbivores](#) (Planktonic Herbivore Temp Dependence)

Contributors	Affiliation	Role
<a href="#">Menden-Deuer, Susanne</a>	University of Rhode Island (URI-GSO)	Principal Investigator
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## Abstract

Quantifying phytoplankton growth as a function of environmental conditions such as temperature is critical for understanding and predicting production in the ocean. Typically, thermal response is described as the steady state growth rates under static conditions. However, here, with a clonal culture of *Heterosigma akashiwo*, temperature was manipulated in the laboratory with the goal of describing how growth rates may change over time through the acclimation process. Growth rates and equivalent spherical diameter of *Heterosigma akashiwo* after temperature transition are reported.

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## Coverage

**Spatial Extent:** Lat:41.65583 Lon:-71.44778

**Temporal Extent:** 2018-06-25 - 2018-09-06

## Dataset Description

Quantifying phytoplankton growth as a function of environmental conditions such as temperature is critical for understanding and predicting production in the ocean. Typically, thermal response is described as the steady state growth rates under static conditions. However, here, with a clonal culture of *Heterosigma akashiwo*, temperature was manipulated in the laboratory with the goal of describing how growth rates may change over time through the acclimation process. Growth rates and equivalent spherical diameter of *Heterosigma akashiwo* after temperature transition are reported.

Also see dataset 'Phytoplankton size over temperature range': <https://www.bco-dmo.org/dataset/783509>

## Methods & Sampling

## Experimental setup

*H. akashiwo* (CCMP 3374) was isolated from Narragansett Bay June 10, 2010 when in situ water temperature was 21.2°C. After isolation the culture was maintained at 15°C. All experiments were conducted with cells that were grown in autoclaved, 0.2 µm sterile-filtered seawater (30-31 ppt) amended with F/2 media without silica (Guillard 1975). All cultures were maintained under a light intensity of 150 µmol photons m<sup>-2</sup> s<sup>-1</sup> and a 12:12 h light: dark cycle. Preliminary experiments to determine the characteristics of this strains' growth patterns were used to maintain cultures in exponential phase by transferring cultures as needed every 4 to 10 days (depending on growth temperature), resulting in cell densities of 500-24,000 cell mL<sup>-1</sup>. To avoid convolution of thermal response and effect of unconditioned media just after transfer culture transfers were restricted from 1 day prior and 1 day post change in temperature (Grabski and Tukaj 2008 ).

## Temperature treatments

Temperature was manipulated in the experiment through a series of sequential temperature shifts. Beginning with a culture which had remained at 15 °C since collection, every four days a triplicate set of the most extreme, current temperature treatments were split with one fraction retained at its current treatment and one fraction shifted a temperature step outward (i.e. further towards the temperature extremes). Only the cultures growing at the highest and lowest temperatures were split and transferred to new temperatures, but all other cultures were continually maintained in exponential growth phase. Cultures maintained at 15 °C throughout the duration of the experiment served as an acclimated control, and as a reference for the temporal consistency in acclimated rates. Including all temperature steps and extrema, the nine treatments included: 6, 8, 10, 12, 18, 22, 25, 28, 31, and the control at 15°C. It took 20 days to complete the individual temperature shifts at 4-day intervals to reach the most extreme temperature treatments.

Dedicated incubators were used for control (15°C, Model 2015 Low Temperature Incubator, VWR Scientific); 4, 6 (I-41LLVL, Percival Scientific); 8, 18, 22 (I-36LLVL, Percival Scientific); and 10 °C (Environ Air, Holman Engineering). Twenty-five, 28, and 31 °C treatments were accomplished with clear 10 L baths controlled by a coupled aquarium heater and thermostat (Fluval, Tru Temp), housed in an illuminated incubator. Light intensity was consistently controlled across temperature treatments.

## Population and growth rate measurements

To quantify changes in cell size, population growth rate, and volumetric growth rate, the abundance and size (Equivalent Spherical Diameter, ESD) distribution of each culture were measured with a 100 µm aperture on a Beckman Coulter Multisizer 3 (Beckman Coulter, Brea California; Kim and Menden-Deuer 2013). The default bin size was the instrument standard of 0.2 µm. Each treatment set was measured daily for 15 days following the initial transfer to the target temperature. Experiments were terminated after 15 days because the objective of these experiments was to establish the response to relatively short-term temperature fluctuations.

## Statistical analysis

Mean and standard deviation of size distribution and cell counts as measured by the Coulter Counter were estimated from the raw data by a gaussian distribution, fit with maximum likelihood estimation to the frequency of particles greater than 8 µm. Measurements that immediately followed culture inoculation and those from the growth were omitted. Cell volumes were estimated with equivalent spherical diameter (ESD) and spherical shape approximation. Cell biomass was calculated using ESD measured with the coulter counter and calculated cell volume with the relationship of pg C cell<sup>-1</sup>=0.288x volume<sup>0.811</sup> (Menden-Deuer and Lessard 2000). Total biomass was approximate using a culture and time specific mean ESD and cell count.

Specific growth rate was calculated by linear regression of natural log transformed abundance. Production rates were calculated by linear regression of natural log transformed total estimated biomass for each culture. Specific growth rates were used to fit a thermal reaction norm modified from Norberg (2004), where *a*, *b*, and *z* are shape parameters, *w* the thermal niche width, *T* a given temperature, and *k(T)* the specific growth rate at that temperature.

$$k(T) = a * e^{bT} [1-(T-z)/w/2)^{-2}]$$

To quantify the effect of time (acclimation) on growth rate the initial growth rates, measured in the first three days ( $\mu_0$ ;  $\Delta \text{time} \leq 3$  days), as well as final rates ( $\mu_f$ ,  $7 \leq \Delta \text{time} \leq 15$  days) were calculated for each temperature treatment.

These experiments were conducted at the University of Rhode Island, Graduate School of Oceanography in the laboratory of Dr. Susanne Menden-Deuer

## Data Processing Description

BCO-DMO Processing Notes:

- added conventional header with dataset name, PI name, version date
- modified parameter names to conform with BCO-DMO naming conventions

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## Data Files

File
<b>growth_rates_ESD.csv</b> (Comma Separated Values (.csv), 1.56 KB) MD5:505018f34fc0ebd45f69647bf87739f6 Primary data file for dataset ID 783500

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## Related Publications

Grabski, K., & Tukaj, Z. (2007). Autoinduction activity of a conditioned medium obtained from high density cultures of the green alga *Scenedesmus subspicatus*. *Journal of Applied Phycology*, 20(3), 323–330.

doi:[10.1007/s10811-007-9260-x](https://doi.org/10.1007/s10811-007-9260-x)

*Methods*

Guillard, R. R. L. (1975). Culture of Phytoplankton for Feeding Marine Invertebrates. *Culture of Marine Invertebrate Animals*, 29–60. doi:[10.1007/978-1-4615-8714-9\\_3](https://doi.org/10.1007/978-1-4615-8714-9_3)

*Methods*

Kim, H., & Menden-Deuer, S. (2013). Reliability of rapid, semi-automated assessment of plankton abundance, biomass, and growth rate estimates: Coulter Counter versus light microscope measurements. *Limnology and Oceanography: Methods*, 11(7), 382–393. doi:[10.4319/lom.2013.11.382](https://doi.org/10.4319/lom.2013.11.382)

*Methods*

Menden-Deuer, S., & Lessard, E. J. (2000). Carbon to volume relationships for dinoflagellates, diatoms, and other protist plankton. *Limnology and Oceanography*, 45(3), 569–579. doi:[10.4319/lo.2000.45.3.0569](https://doi.org/10.4319/lo.2000.45.3.0569)

*Methods*

Norberg, J. (2004). Biodiversity and ecosystem functioning: A complex adaptive systems approach. *Limnology and Oceanography*, 49(4,part2), 1269–1277. doi:[10.4319/lo.2004.49.4\\_part\\_2.1269](https://doi.org/10.4319/lo.2004.49.4_part_2.1269)

*Methods*

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## Parameters

Parameter	Description	Units
Culture	culture identifier	unitless
Time	either growth rate immediately after temperature change (t1) or growth rate after 1 week to acclimate (tf)	unitless
Temperature	temperature treatment	degrees Celsius
GrowthRate	divison-based growth rate	per day
BioRate	biovolume-based growth rate	per day
MeanESD_t	Mean Equivalent Spherical Diameter for a culture during the period of growth rate calculation	micrometers

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## Instruments

<b>Dataset-specific Instrument Name</b>	Beckman Coulter Multisizer III Counter
<b>Generic Instrument Name</b>	Coulter Counter
<b>Dataset-specific Description</b>	Used to count cells.
<b>Generic Instrument Description</b>	An apparatus for counting and sizing particles suspended in electrolytes. It is used for cells, bacteria, prokaryotic cells and virus particles. A typical Coulter counter has one or more microchannels that separate two chambers containing electrolyte solutions. from <a href="https://en.wikipedia.org/wiki/Coulter_counter">https://en.wikipedia.org/wiki/Coulter_counter</a>

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## Project Information

### Quantifying Temperature Dependence In Growth & Grazing Rates of Planktonic Herbivores (Planktonic Herbivore Temp Dependence)

**Coverage:** Narragansett Bay

#### *NSF Award Abstract:*

Plankton, single-celled organisms that inhabit the world's oceans are responsible for the generation of oxygen, cycling energy and matter between the atmosphere and the deep ocean and are the basis for virtually all seafood harvested. These life-giving functions critically depend on the relative rates at which plankton grow

and get eaten. How temperature influences those rates is essential to understand plankton responses to environmental changes and ocean dynamics. It is well established that plankton grow faster when temperatures are higher however, whether feeding has a similar temperature dependence is unknown. That means oceanographers are missing key data required to build global predictive models. This project will fill essential knowledge gaps and measure physiological rates of singled celled zooplankton across temperature gradients representing the global ocean, from polar to tropical regions and throughout the seasonal cycle. Researchers will combine laboratory experiments with specimens taken from the coastal ocean (Narragansett Bay), which is exemplary in its strong seasonal temperature variations. These data will provide a clear picture of the production capacity and activity of plankton in a global and dynamic ocean. The project supports an early career scientist, as well as graduate and undergraduate students. Scientists will continue communicating their research to the public through large-scale outreach events, education at the high-school level, and engagement through online and other media. Moreover, researchers will continue collaborating with the Metcalf Institute for Marine & Environmental Reporting to support their Annual Science Immersion Workshop for Journalists and their ongoing work to disseminate research findings through web-based seminars.

Grazing is the single largest loss factor of marine primary production and thus affects a key transfer rate between global organic and inorganic matter pools. Remarkably, data for herbivorous protist growth and grazing rates at temperatures representative of the vast polar regions and during winter and spring periods are extremely sparse. By combining laboratory experiments with ground truthing fieldwork, this project alleviates a central knowledge gap in oceanography and delivers the empirical measurements necessary to derive algorithms to incorporate temperature dependence of heterotrophic protist growth and grazing rates into biogeochemical models. The extraordinary seasonal temperature fluctuations in a temperate coastal estuary (Narragansett Bay) are exploited to measure rates of heterotrophic protists isolated from different temperatures and seasons and to quantify the temperature and acclimation responses of these ecotypes. This project delivers data urgently needed to solve the conundrum of whether herbivorous growth and predation is depressed at low temperatures, implying low trophic transfer rates and high carbon export, or if predation proceeds at rates comparable to temperate systems with primary production largely lost to predation. Large temperature gradients in the global ocean mean that cross-biome and biogeochemical models are particularly sensitive to assumptions about the temperature dependence in modeled rate processes. Establishment of the dependence of heterotrophic plankton physiological rates (growth and grazing) to gradients of temperature, mimicking realistic conditions experienced by plankton in a changing ocean, is a key step towards integrating much needed biological information in biogeochemical modeling efforts. This project makes a significant contribution to linking ecological research with ecosystem models by providing empirically rooted algorithms of the temperature dependence of protistan herbivory and growth rates, key processes in the transformation of organic matter in global biogeochemical cycles and tools critically missing in ecosystem models.

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## Funding

Funding Source	Award
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-1736635</a>

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