Microbial 16S OTU count data from staghorn coral health experiments with treatment with and without antibiotics

Website: https://www.bco-dmo.org/dataset/816379
Data Type: experimental
Version: 1
Version Date: 2020-06-25

Project
» Coral-microbial interactions as determinants of disease dynamics (Coral-microbial interactions)

Contributors	Affiliation	Role
Gouhier, Tarik C.	Northeastern University	Principal Investigator
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Abstract
This dataset reports the number of reads of microbial 16S OTU's from a priority experiment with staghorn coral treated with and without antibiotics.

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Coverage

Spatial Extent: **Lat**:9.3513 **Lon**:-82.2565
Temporal Extent: 2015-06 - 2015-06

Dataset Description

This dataset reports the number of reads of microbial 16S OTU's from a priority experiment with staghorn coral treated with and without antibiotics. Please note that this dataset contains over 1.4 million records and is slow to load on the data display page. The data file is available for download via the "Data Files" section.

Methods & Sampling

A priority experiment was conducted in 24 recirculating tanks at the Smithsonian stations in Bocas del Toro, Panama. Replicate staghorn corals fragments from 10 genotypes were placed into the tanks containing 12 liters of UV sterilized seawater. Twelve tanks were then exposed to an antibiotic cocktail of Kanamycin, Ampicillan, Tetracycline and Choloramphenicol (100mg/ml each) twice over a 48hr period, and the corals from the antibiotic treated and untreated tanks were sampled for DNA (T0 sample point) and lesioned with an airbrush. Half of the antibiotic- treated and untreated tanks were then exposed to 30ml of disease slurry (D) or healthy slurry (H) for the first exposure. 24hrs later a second dose of disease or healthy slurry was applied to complete the priority experiment treatments of healthy_healthy (h_h), disease_disease (d_d), healthy_disease

(h_d), and disease_healthy (d_h). DNA for the microbial analyses were sampled 18hrs later (T2) and then disease was monitored over the course of the experiment. Corals that developed disease were sampled (T3) and sacrificed. All of the corals were sampled (T3 timepoint) on day 7.

DNA from preserved coral samples were extracted using an Agencourt DNAdvance kit, in order to prepare 16S rDNA profiling libraries. 16s libraries of the hypervariable V3_v4 region were then prepared using a two-step PCR protocol and combinatorial barcodes, resulting in amplicon lengths from ~300-400 bp paired-end joined sequences from Illumina 250 bp sequencing on an Illumina HiSeq 2500.

16s sequencing data were clustered into operational taxonomic units (OTUs) using a 97% identity threshold in QIIME. Default QIIME settings were used to align reads, to remove chimera sequences, and to assign taxonomy.

Data Processing Description

BCO-DMO Processing Notes:
- added conventional header with dataset name, PI name, version date
- converted color codes to full color words
- removed column 'tank'

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Data Files

File	
Microbe priority experiment: OTU counts filename: microbes_counts.csv	(Comma Separated Values (.csv), 37.35 MB) MD5:c2ae56e29ddf4fc4d680d8d6a5545e4a
Microbial 16S OTU count data from staghorn coral health experiments with treatment with and without antibiotics. PI's: T. Gouhier, S. Vollmer (NEU); version date: 2020-06-25	
microbes_counts.csv	(Comma Separated Values (.csv), 37.35 MB) MD5:c2ae56e29ddf4fc4d680d8d6a5545e4a
Primary data file for dataset ID 816379	

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Related Publications

Chu, N. D., & Vollmer, S. V. (2016). Caribbean corals house shared and host-specific microbial symbionts over time and space. Environmental Microbiology Reports, 8(4), 493–500. doi:10.1111/1758-2229.12412
Methods

Dunphy, C. M., Gouhier, T. C., Chu, N. D., & Vollmer, S. V. (2019). Structure and stability of the coral microbiome in space and time. Scientific Reports, 9(1). doi:10.1038/s41598-019-43268-6
Methods

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Related Datasets

IsRelatedTo

Gouhier, T. C., Vollmer, S. V. (2021) **Microbial 16S OTU annotation information from staghorn coral health experiments with treatment with and without antibiotics.** Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2020-06-26 doi:10.26008/1912/bco-dmo.816367.1 [view at BCO-DMO]

Gouhier, T. C., Vollmer, S. V. (2021) **Staghorn coral health following experimental treatment with and without antibiotics.** Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2020-06-25 doi:10.26008/1912/bco-dmo.816316.1 [view at BCO-DMO]

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Parameters

Parameter	Description	Units
OTUname	OTU ID/label	unitless
genotype	Coral host genotype with levels represented by labels corresponding to different colors	unitless
antibiotic	Experimental antibiotic treatment: levels PlusA or MinusA	unitless
exposure	Two experimental exposures to microbes from healthy or diseased corals: levels Hh (healthy then healthy); Hd (healthy then disease); Dh (disease then healthy); Dd (disease then disease)	unitless
time	Time points since antibiotic treatment was administered: T0; T2; T3	hours
count	OTU counts	OTU's

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Instruments

Dataset-specific Instrument Name	Illumina HiSeq 2500
Generic Instrument Name	Automated DNA Sequencer
Generic Instrument Description	General term for a laboratory instrument used for deciphering the order of bases in a strand of DNA. Sanger sequencers detect fluorescence from different dyes that are used to identify the A, C, G, and T extension reactions. Contemporary or Pyrosequencer methods are based on detecting the activity of DNA polymerase (a DNA synthesizing enzyme) with another chemoluminescent enzyme. Essentially, the method allows sequencing of a single strand of DNA by synthesizing the complementary strand along it, one base pair at a time, and detecting which base was actually added at each step.

Dataset-specific Instrument Name	
Generic Instrument Name	Thermal Cycler
Generic Instrument Description	A thermal cycler or "thermocycler" is a general term for a type of laboratory apparatus, commonly used for performing polymerase chain reaction (PCR), that is capable of repeatedly altering and maintaining specific temperatures for defined periods of time. The device has a thermal block with holes where tubes with the PCR reaction mixtures can be inserted. The cycler then raises and lowers the temperature of the block in discrete, pre-programmed steps. They can also be used to facilitate other temperature-sensitive reactions, including restriction enzyme digestion or rapid diagnostics. (adapted from http://serc.carleton.edu/microbelife/research_methods/genomics/pcr.html)

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Project Information

Coral-microbial interactions as determinants of disease dynamics (Coral-microbial interactions)

Coverage: Panama, Bocas del Toro Archipelago

Description from NSF award abstract:
The health of numerous animal and plant hosts depends on the composition of their microbiome. Although the health of hosts is often linked to environmental factors that favor the growth of pathogenic microbes, the diversity of animal and plant microbiomes suggests that competition plays an important role in determining the frequency and severity of disease outbreaks. This project uses the endangered Caribbean staghorn coral as a model system to understand how the environment and microbial interactions controls the spread of White Band Disease (WBD), a bacterial disease that has decimated nearly 95% of Caribbean staghorn coral populations. A combination of manipulative experiments, field surveys and mathematical modeling will be used to determine how water temperature, microbial movement between- and microbial competition within coral hosts influence WBD incidence. By identifying the environmental and microbial drivers of WBD, this project will allow managers to (i) predict hotspots of vulnerability to WBD in space and time, and (ii) identify optimal strategies for restoring these once prominent members of Caribbean coral reef communities. This research will address important societal needs by cross-training graduate students in coral biology, microbial genetics, bioinformatics, mathematical modeling, computer programming and statistics. Results of this project will be integrated into undergraduate courses in genetics, ecological modeling and biostatistics in order to emphasize the importance of quantitative and interdisciplinary STEM training for addressing important questions in biology. Finally, a series of interactive web modules will be created to disseminate the results of this project beyond academic circles, including to Northeastern University's Marine Science Center K-12 outreach programs and the Smithsonian Tropical Research Institute's outreach programs in Panama.

There is growing recognition that the processes that structure microbial communities may scale up to explain disease outbreaks in their hosts. Despite the complexity of microbial communities, most studies to date have focused on resolving the direct relationship between the environment, the occurrence of pathogenic microbes, and the incidence of disease. However, the effects of microbial species interactions and dispersal on the emergence of host diseases remain largely unknown. This project will combine microbial genetics and mathematical modeling to understand the relative influence of the environment, species interactions and dispersal on the structure of microbial communities and the dynamics of disease in their coral hosts. This research uses the endangered Caribbean staghorn coral (Acropora cervicornis) and White Band Disease (WBD) as a model host-pathogen system. This once dominant, reef-building coral was decimated by WBD, prompting its listing as an endangered species. Recent work in this system suggests that (i) bacteria are the cause of WBD, (ii) the microbial community living within the host can produce antibiotic compounds that suppress pathogenic bacteria, and (iii) temperature increase promotes infection and reduces the production of antibiotic compounds. These findings suggest that the interplay between the environment and host-associated microbial species determines the structure of the microbial community and the health of the coral host. To disentangle these processes, a multi-factorial transmission experiment will be conducted to understand the direct and indirect effects of temperature, pathogen exposure, and microbial community complexity on disease dynamics. To determine how these results scale up to natural coral reefs, a spatial coral-microbial model will be fitted to field survey data. This fitted model will elucidate how seasonal temperature variation and microbial dispersal jointly influence coral disease outbreaks and the structure of coral-microbial communities across spatial scales. The proposed research will integrate research with teaching and training of undergraduate students.

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Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-1458158

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