

Seawater chemistry and benthic metabolic rates as a function of coral cover from a mesocosm experiment conducted at the Hawaii Institute of Marine Biology in June 2016

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Data Type: Other Field Results, experimental

Version: 1

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Project

» [CAREER: Biogeochemical Modification of Seawater CO₂ Chemistry in Near-Shore Environments: Effect of Ocean Acidification](#) (Nearshore CO₂)

Contributors	Affiliation	Role
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Abstract

Seawater chemistry and benthic metabolic rates as a function of coral cover from a mesocosm experiment conducted at the Hawaii Institute of Marine Biology in June 2016.

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Coverage

Spatial Extent: Lat:21.433019 Lon:-157.786461

Temporal Extent: 2016-06-14 - 2016-06-17

Methods & Sampling

General study design:

Coral communities with different benthic cover (0, 40, 80%) were created in flow-through mesocosms exposed to natural environmental variations. The experiment was designed to examine how benthic metabolism influence diurnal seawater carbonate chemistry and how this scales with varying coral cover. The experiment was conducted at the Coral Reef Ecology Laboratory at the Hawaii Institute of Marine Biology in Kaneohe, HI, USA.

Methods description:

Coral communities with planar surface area coverage ranging 0, 20, 40, 60, 80 and 100 percent were created within flow-through mesocosms. Diurnal seawater chemistry was measured for communities during June 2016 by taking hourly water measurements (T, S, DO_{sat}, DO_{conc}, and pH) and collecting water samples for dissolved inorganic carbon (DIC) and total alkalinity (TA) every 3 hours over 24-hour time periods. Seawater chemistry data were used to calculate diurnal net community production (NCP) and net community calcification (NCC) rates using modified standard equations (Langdon et al. 2010).

Analytical Methods:

Seawater samples were collected by hand using 250 ml Pyrex glass bottles and fixed with 100 µL HgCl₂ as per standard protocols (Dickson et al. 2007). Handheld YSI multi-meter instrument was used to measure temperature, salinity, dissolved oxygen, and pH at the time of sampling. All seawater samples were transported to the Scripps Coastal and Open Ocean Biogeochemistry lab and analyzed for TA via an open-cell potentiometric acid titration system developed at Scripps Institution of Oceanography (SIO) by A. Dickson (Dickson et al. 2007) and DIC via an automated infra-red inorganic carbon analyzer (AIRICA, Marianda Inc).

Quality Control:

Standard protocols were followed for sampling and analysis of seawater TA and DIC (Dickson et al. 2007). YSI multi-meter instrument was calibrated prior to each sampling period with accuracies of 0.2 °C for temperature, 1% for salinity, ±2% for dissolved oxygen saturation, 0.2 mg/L for dissolved oxygen concentration, and 0.2 for pH. The accuracy and precision of TA (0.33 ± 2.21 µmol/kg) and DIC (-1.31 ± 2.98 µmol/kg) measurements were evaluated using certified reference materials (CRM) provided by the laboratory of A. Dickson at SIO and analyzed every 5 samples for DIC and ~15-20 samples for TA.

For additional information please see Page et al. (2017).

Known Problems:

Rows for Mesocosm_IDs of "Header 1", "9", and "10" on Date 14-06-16 and Time 00:00 (lines 56, 60, 62 in original Excel file): TA did not pass QAQC and therefore were omitted. Consequently, NCC and NCP could not be calculated for these data.

Data Processing Description

Data Processing:

Carbonate chemistry (pH_calc, pCO₂_calc, and omega_calc) was constrained using CO₂SYS software (Lewis and Wallace 1998). Data were processed and statistically compared using R Software (v 3.2.0).

BCO-DMO Processing:

- added UTC date/time field in ISO8601 format.

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Related Publications

Page, H. N., Courtney, T. A., Collins, A., De Carlo, E. H., & Andersson, A. J. (2017). Net Community Metabolism and Seawater Carbonate Chemistry Scale Non-intuitively with Coral Cover. *Frontiers in Marine Science*, 4. doi:[10.3389/fmars.2017.00161](https://doi.org/10.3389/fmars.2017.00161)

Results

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Parameters

Parameter	Description	Units
Date	Date of measurements (local time; HST); format: DD-MM-YY	unitless
Time	Time of measurements (local time; HST); format: hh:mm	unitless
ISO_DateTime_UTC	Date and time of measurements (UTC) in ISO8601 format: YYYY-MM-DDThh:mmZ	unitless
MesocosmID	Mesocosm identification	unitless
Treatment	Each mesocosm acted as inflow or contained a coral community. 0=0% coral cover; 20 = 20% coral cover; 40 = 40% coral cover, 60 = 60% coral cover, 80 = 80% coral cover, and 100 = 100% coral cover.	unitless
Temp	Temperature	degrees Celsius
Salinity	Salinity	unitless
DO_sat	dissolved oxygen percent saturation	unitless (percent)
DO_conc	dissolved oxygen concentration	milligrams per liter (mg/L)
pH_ysi	pH	unitless
DIC	dissolved inorganic carbon	micromoles per kilogram (umol/Kg)
TA	total alkalinity	micromoles per kilogram (umol/Kg)
pH_calc	pH (total scale) calculated from in situ temperature, salinity, TA, and DIC	unitless
pCO2_calc	pCO2 calculated from in situ temperature, salinity, TA, and DIC	microatmospheres (uatm)
omega_calc	Seawater saturation state with respect to aragonite calculated from in situ temperature, salinity, TA, and DIC	unitless
NCC	Net community calcification rate	millimoles per square meter per hour (mmol/m ² /h)
NCP	Net community primary production rate	millimoles per square meter per hour (mmol/m ² /h)

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Instruments

Dataset-specific Instrument Name	Open-cell potentiometric acid titration system
Generic Instrument Name	Automatic titrator
Dataset-specific Description	The open-cell potentiometric acid titration system was developed by the laboratory of A.G. Dickson. Briefly, a known amount of seawater is added to an open cell beaker where hydrochloric acid as acidified to a pH of 3.5-4.0 and allowed to stabilize to remove CO ₂ gas formed by the addition of acid. Small aliquots of hydrochloric acid are then added to pH of ~ 3.0. The titration is monitored by a glass electrode and the total alkalinity of the sample is calculated using a non-linear least-squares method following Dickson et al. (2007).
Generic Instrument Description	Instruments that incrementally add quantified aliquots of a reagent to a sample until the end-point of a chemical reaction is reached.

Dataset-specific Instrument Name	AIRICA (Marianda Inc.)
Generic Instrument Name	Inorganic Carbon Analyzer
Dataset-specific Description	The Automated Infra Red Inorganic Carbon Analyzer (AIRICA) utilizes infrared detection of CO ₂ gas purged from an acidified seawater sample. A high-precision syringe pump extracts the seawater sample, acidifies the sample with phosphoric acid, and analyzes the gas released with an infrared light analyzer (LICOR). The CO ₂ signal is integrated for each sample to quantify the total inorganic carbon for a given aliquot of seawater analyzed. Three aliquots and peak integrations are performed for each seawater sample and averaged to determine the dissolved inorganic carbon for each sample. Please see http://marianda.com/index.php?site=products&subsite=airica for a complete instrument description.
Generic Instrument Description	Instruments measuring carbonate in sediments and inorganic carbon (including DIC) in the water column.

Dataset-specific Instrument Name	YSI Professional Plus (Pro Plus) Handheld Multiparameter
Generic Instrument Name	Water Quality Multiprobe
Dataset-specific Description	YSI Handheld Multiparameter Instruments were used to monitor in situ temperature (accuracy $\pm 0.2^{\circ}\text{C}$), salinity (accuracy $\pm 1\%$), DO _{mg} (accuracy ± 0.2 mg/L), and DO _% (accuracy $\pm 2\%$), and pH (accuracy ± 0.2).
Generic Instrument Description	An instrument which measures multiple water quality parameters based on the sensor configuration.

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Project Information

CAREER: Biogeochemical Modification of Seawater CO₂ Chemistry in Near-Shore Environments:

Effect of Ocean Acidification (Nearshore CO2)

Coverage: San Diego, California; Bermuda; Oahu, Hawaii

NSF Award Abstract:

Because of well-known chemical principles, changes in the CO₂ chemistry of seawater in the open ocean as a result of rising atmospheric CO₂ can be predicted very accurately. On the other hand, in near-shore environments, these projections are much more difficult because the CO₂ chemistry is largely modified by biogeochemical processes operating on timescales of hours to months. To make predictions on how near-shore seawater CO₂ chemistry will change in response to ocean acidification (OA), it is critical to consider the relative influence of net ecosystem production (NEP) and net ecosystem calcification (NEC), and how these processes might change in response to this major perturbation. Understanding how future OA will alter near-shore seawater CO₂ chemistry and variability was identified as a major critical knowledge gap at the recent IPCC WG II/WG I workshop on impacts of ocean acidification on marine biology and ecosystems in January of 2011, and also at the International Ocean Acidification Network workshop in Seattle in June of 2012.

With funding from this CAREER award, a researcher at the Scripps Institute of Oceanography and his students will study how biogeochemical processes and the relative contributions from NEP and NEC modify seawater CO₂ chemistry in near-shore environments influenced by different benthic communities under well-characterized environmental and physical conditions, and how these processes might change in response to OA. The team will investigate a limited number of contrasting habitats in subtropical (reef crest, back/patch reef, lagoon, seagrass bed, algal mat) and temperate (kelp bed, inter- and sub-tidal, marsh) environments during summer and winter, employing a method that evaluates the function and performance of the carbon cycle of a system using a stoichiometric vector approach based on changes in total dissolved inorganic carbon (DIC) and total alkalinity (TA). These field studies will be complemented by controlled mesocosm experiments with contrasting and mixed benthic communities under different OA scenarios.

The project has two educational components: (1) developing a research-driven OA and biogeochemistry course based on inquiry-, experience-, and collaborative-based learning; and (2) working with the Ocean Discovery Institute (ODI) to engage individuals from a local underrepresented minority community in science through educational activities focused on OA, and also providing a moderate number of internships for high school and college students to engage in this research project.

Broader Impacts: This project will directly support one PhD student, one junior research technician, and two high school and college interns from underrepresented minorities (URM) each summer of the project. It will contribute to the education of 80 undergraduate and graduate students participating in the research based ocean acidification/biogeochemistry course offered four times throughout the duration of the project at SIO/UCSD. Education and curricular material on the topics of OA, including hands-on laboratories, classroom and field-based activities will be developed through the collaboration with the ODI and brought to hundreds of URM students and their teachers in the City Heights area, a community with the highest poverty and ethnic diversity in the San Diego region. This collaboration will enable URM students to directly engage in a rapidly evolving field of research that has high relevance at both the local and global scales. To ensure broad dissemination of this project and the topic of OA, the research team will work with the Google Ocean team to incorporate information and educational material in the Google Ocean Explorer.

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Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-1255042

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