# Biomass of taxonomic groups from manually corrected live Imaging FlowCytobot (IFCB) analysis of water samples collected from surface and chlorophyll maximum depths during R/V Pt. Sur cruise PS 18-09 in the western Gulf of Mexico, Sept-Oct 2017

Website: https://www.bco-dmo.org/dataset/840147

**Data Type**: Cruise Results

Version: 1

Version Date: 2021-02-08

#### **Project**

» RAPID: Hurricane Impact on Phytoplankton Community Dynamics and Metabolic Response (HRR)

| Contributors        | Affiliation   | Role                      |
|---------------------|---|---------------------------|
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#### Abstract

Biomass data for taxonomic groups from manually corrected live Imaging FlowCytobot (IFCB) analysis of water samples collected from surface and chlorophyll maximum depths during R/V Pt. Sur PS 18-09, western Gulf of Mexico, Sept-Oct 2017. These data were inspected visually and manually corrected.

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# Coverage

**Spatial Extent**: N:29.0649 E:-94.9 S:27.2286 W:-97.268

**Temporal Extent**: 2017-09-23 - 2017-10-01

## Methods & Sampling

On each of 2 cruise legs 01 and 03, samples were collected at 7 stations (S01, S06, S11, S16, S21, SS and GI) from 2 depths [surface and chlorophyll maximum depth when possible; see HRR-bottle data]) by CTD-rosette. At each station, triplicate 5-ml samples pre-filtered through 150  $\mu$ m Nitex were analyzed immediately with an onboard Imaging FlowCytobot. All image data can be viewed on the TOAST dashboard: https://toast.tamu.edu/timeline?dataset=HRR cruise.

Image analysis and feature extraction were performed using software developed by Sosik and colleagues

which is available on github (<a href="https://github.com/hsosik/ifcb-analysis/">https://github.com/hsosik/ifcb-analysis/</a>). The automated classification approach of Sosik & Olson (2007), as modified and described by Anglès et al. (2019), was employed and the automated classification results were then inspected visually and **manually** corrected into a total of 102 categories that included 35 categories of diatoms, 30 categories of dinoflagellates, 10 categories of ciliates, 10 categories of flagellates, and 17 'others', which included filamentous cyanobacteria, freshwater chlorophytes, coccolithophorids, and small cells that could not be identified taxonomically from images (refer to Fiorendino et al. 2021. for more details). See related dataset.

For comparison with the Texas Observatory for Algal Succession Time series (TOAST), IFCB images were also classified automatically into one of 112 classes utilizing a custom convolutional neural network (**CNN**) trained on a curated set of images (Henrichs et al. 2021.). See related dataset.

**This dataset: Biomass** for each image was estimated using the algorithm developed by Moberg & Sosik (2012) to calculate cellular volume from the extracted image features and then convert to total carbon per image (Menden-Deuer & Lessard 2000) and summed for each class..

#### Sampling locations:

| Sample ID | Station | Leg | Location<br>Lat <sup>o</sup> N/Long <sup>o</sup> W |
|-----------|---------|-----|--|
| L1_S01    | S01     | 1   | 27 2206 07 2606                                    |
| L3_S01    | S01     | 3   | 27.2286 -97.2686                                   |
| L1_S06    | S06     | 1   | 27.8358 -96.9874                                   |
| L3_S06    | S06     | 3   | 27.0330 -90.9074                                   |
| L1_S11    | S11     | 1   | 28.2614 -96.4129                                   |
| L3_S11    | S11     | 3   | 20.2014 -90.4129                                   |
| L1_S16    | S16     | 1   | 28.5366 -95.8656                                   |
| L3_S16    | S16     | 3   | 20.5500 -95.0050                                   |
| L1_S21    | S21     | 1   | 28.7644 -95.2978                                   |
| L3_S21    | S21     | 3   | 20.7044 -33.2370                                   |
| L1_SS     | SS      | 1   | 28.9600 -95.0946                                   |
| L3_SS     | SS      | 3   | 20.9000 -95.0940                                   |
| L1_GI     | GI      | 1   | 29.0649 -94.9000                                   |
| L3_GI     | GI      | 3   |  |

#### **Data Processing Description**

#### - BCO-DMO Processing Notes:

- data submitted in Excel file "manual carbon ifcb.xlsx" sheet "Biomass" extracted to csv
- added conventional header with dataset name, PI name, version date
- modified parameter names to conform with BCO-DMO naming conventions
- rounded values to 2 decimal places

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# **Data Files**

#### File

IFCB\_manual\_carbon.csv(Comma Separated Values (.csv), 14.91 KB)

MD5:600d135449c74be4238709b917d6928d

Primary data file for dataset ID 840147

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## **Related Publications**

Anglès, S., Jordi, A., Henrichs, D. W., & Campbell, L. (2019). Influence of coastal upwelling and river discharge on the phytoplankton community composition in the northwestern Gulf of Mexico. Progress in Oceanography, 173, 26–36. doi:10.1016/j.pocean.2019.02.001

Results

Fiorendino, J. M., Gaonkar, C. C., Henrichs, D. W., & Campbell, L. (2021). Drivers of microplankton community assemblage following tropical cyclones. Journal of Plankton Research, 45(1), 205–220. https://doi.org/10.1093/plankt/fbab073

Results

Henrichs, D. W., Anglès, S., Gaonkar, C. C., & Campbell, L. (2021). Application of a convolutional neural network to improve automated early warning of harmful algal blooms. Environmental science and pollution research international, 28(22), 28544–28555. https://doi.org/10.1007/s11356-021-12471-2

Results

Menden-Deuer, S., & Lessard, E. J. (2000). Carbon to volume relationships for dinoflagellates, diatoms, and other protist plankton. Limnology and Oceanography, 45(3), 569–579. doi:10.4319/lo.2000.45.3.0569

Methods

Moberg, E. A., & Sosik, H. M. (2012). Distance maps to estimate cell volume from two-dimensional plankton images. Limnology and Oceanography: Methods, 10(4), 278–288. doi: 10.4319/lom.2012.10.278

Methods

Olson, R. J., & Sosik, H. M. (2007). A submersible imaging-in-flow instrument to analyze nano-and microplankton: Imaging FlowCytobot. Limnology and Oceanography: Methods, 5(6), 195–203. doi:10.4319/lom.2007.5.195

Methods

Sosik, H. M., & Olson, R. J. (2007). Automated taxonomic classification of phytoplankton sampled with imaging-in-flow cytometry. Limnology and Oceanography: Methods, 5(6), 204–216. doi:10.4319/lom.2007.5.204

Methods

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## **Related Datasets**

#### **IsRelatedTo**

Imaging FlowCytobot (IFCB) analysis of water samples collected from surface and chlorophyll maximum depths during R/V Pt. Sur cruise PS 18-09 in the western Gulf of Mexico, Sept-Oct 2017. Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2021-02-08 doi:10.26008/1912/bco-dmo.840147.1 [view at BCO-DMO]

Campbell, L., Henrichs, D. W. (2021) **Cell abundances for taxonomic groups from manually corrected live Imaging FlowCytobot (IFCB) analysis of water samples collected from surface and chlorophyll maximum depths during R/V Pt. Sur cruise PS 18-09 in the western Gulf of Mexico, Sept-Oct 2017.** Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2021-02-08 doi:10.26008/1912/bco-dmo.840060.1 [view at BCO-DMO]

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#### **Parameters**

| Parameter      | Description  | Units                      |
|----------------|--|----------------------------|
| Major_Category | 5 major groups of microplankton  | unitless                   |
| Class          | Taxonomic names of the species or genus (if known) or group identified | unitless                   |
| L1GI_1         | Biomass of class at Leg 1 station GI; replicate 1                      | micrograms<br>carbon/liter |
| L1GI_2         | Biomass of class at Leg 1 station GI; replicate 2                      | micrograms<br>carbon/liter |
| L1GI_3         | Biomass of class at Leg 1 station GI; replicate 3                      | micrograms<br>carbon/liter |
| L1SS_1         | Biomass of class at Leg 1 station SS; replicate 1                      | micrograms<br>carbon/liter |
| L1SS_2         | Biomass of class at Leg 1 station SS; replicate 2                      | micrograms<br>carbon/liter |
| L1SS_3         | Biomass of class at Leg 1 station SS; replicate 3                      | micrograms<br>carbon/liter |
| L1S21_1        | Biomass of class at Leg 1 station S21; replicate 1                     | micrograms<br>carbon/liter |
| L1S21_2        | Biomass of class at Leg 1 station S21; replicate 2                     | micrograms<br>carbon/liter |
| L1S21_3        | Biomass of class at Leg 1 station S21; replicate 3                     | micrograms<br>carbon/liter |
| L1S16_1        | Biomass of class at Leg 1 station S16; replicate 1                     | micrograms<br>carbon/liter |
| L1S16_2        | Biomass of class at Leg 1 station S16; replicate 2                     | micrograms<br>carbon/liter |
| L1S16_3        | Biomass of class at Leg 1 station S16; replicate 3                     | micrograms<br>carbon/liter |
| L1S11_1        | Biomass of class at Leg 1 station S11; replicate 1                     | micrograms<br>carbon/liter |
| L1S11_2        | Biomass of class at Leg 1 station S11; replicate 2                     | micrograms<br>carbon/liter |
| L1S11_3        | Biomass of class at Leg 1 station S11; replicate 3                     | micrograms<br>carbon/liter |
| L1S06_1        | Biomass of class at Leg 1 station S06; replicate 1                     | micrograms<br>carbon/liter |

| L1S06_2 | Biomass of class at Leg 1 station S06; replicate 2 | micrograms<br>carbon/liter |
|---------|--|----------------------------|
| L1S06_3 | Biomass of class at Leg 1 station S06; replicate 3 | micrograms<br>carbon/liter |
| L1S01_1 | Biomass of class at Leg 1 station S01; replicate 1 | micrograms<br>carbon/liter |
| L1S01_2 | Biomass of class at Leg 1 station S01; replicate 2 | micrograms<br>carbon/liter |
| L1S01_3 | Biomass of class at Leg 1 station S01; replicate 3 | micrograms<br>carbon/liter |
| L3GI_1  | Biomass of class at Leg 3 station GI; replicate 1  | micrograms<br>carbon/liter |
| L3GI_2  | Biomass of class at Leg 3 station GI; replicate 2  | micrograms<br>carbon/liter |
| L3GI_3  | Biomass of class at Leg 3 station GI; replicate 3  | micrograms<br>carbon/liter |
| L3SS_1  | Biomass of class at Leg 3 station SS; replicate 1  | micrograms<br>carbon/liter |
| L3SS_2  | Biomass of class at Leg 3 station SS; replicate 2  | micrograms<br>carbon/liter |
| L3SS_3  | Biomass of class at Leg 3 station SS; replicate 3  | micrograms<br>carbon/liter |
| L3S21_1 | Biomass of class at Leg 3 station S21; replicate 1 | micrograms<br>carbon/liter |
| L3S21_2 | Biomass of class at Leg 3 station S21; replicate 2 | micrograms<br>carbon/liter |
| L3S21_3 | Biomass of class at Leg 3 station S21; replicate 3 | micrograms<br>carbon/liter |
| L3S16_1 | Biomass of class at Leg 3 station S16; replicate 1 | micrograms<br>carbon/liter |
| L3S16_2 | Biomass of class at Leg 3 station S16; replicate 2 | micrograms<br>carbon/liter |
| L3S16_3 | Biomass of class at Leg 3 station S16; replicate 3 | micrograms<br>carbon/liter |
| L3S11_1 | Biomass of class at Leg 3 station S11; replicate 1 | micrograms<br>carbon/liter |
| L3S11_2 | Biomass of class at Leg 3 station S11; replicate 2 | micrograms<br>carbon/liter |
| L3S11_3 | Biomass of class at Leg 3 station S11; replicate 3 | micrograms<br>carbon/liter |
| L3S06_1 | Biomass of class at Leg 3 station S06; replicate 1 | micrograms<br>carbon/liter |
| L3S06_2 | Biomass of class at Leg 3 station S06; replicate 2 | micrograms<br>carbon/liter |
| L3S06_3 | Biomass of class at Leg 3 station S06; replicate 3 | micrograms<br>carbon/liter |
| L3S01   | Biomass of class at Leg 3 station S01              | micrograms<br>carbon/liter |

# **Instruments**

| Dataset-<br>specific<br>Instrument<br>Name | Image FlowCytobot (McLane Research Laboratories, Inc.)   |  |
|--|--|--|
| Generic<br>Instrument<br>Name              | Imaging FlowCytobot  |  |
| Generic<br>Instrument<br>Description       | The Imaging FlowCytobot (IFCB) is an in-situ automated submersible imaging flow cytometer that generates images of particles in-flow taken from the aquatic environment. <a href="https://mclanelabs.com/imaging-flowcytobot/">https://mclanelabs.com/imaging-flowcytobot/</a> |  |

| Dataset-<br>specific<br>Instrument<br>Name |   |
|--|---|
| Generic<br>Instrument<br>Name              | Niskin bottle   |
| Dataset-<br>specific<br>Description        | Used to collect samples   |
| Description                                | A Niskin bottle (a next generation water sampler based on the Nansen bottle) is a cylindrical, non-metallic water collection device with stoppers at both ends. The bottles can be attached individually on a hydrowire or deployed in 12, 24, or 36 bottle Rosette systems mounted on a frame and combined with a CTD. Niskin bottles are used to collect discrete water samples for a range of measurements including pigments, nutrients, plankton, etc. |

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# **Deployments**

# PS1809

| Website     | https://www.bco-dmo.org/deployment/784313  |
|-------------|--|
| Platform    | R/V Point Sur  |
| Start Date  | 2017-09-22   |
| End Date    | 2017-10-03   |
| Description | HRR study with three legs. Chief Scientists: Steve DiMarco (Leg 1); Kristen Thyng (Leg 2); Lisa Campbell (Leg 3). R2R Cruise Page: <a href="https://www.rvdata.us/search/cruise/PS1809">https://www.rvdata.us/search/cruise/PS1809</a> |

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# **Project Information**

RAPID: Hurricane Impact on Phytoplankton Community Dynamics and Metabolic Response (HRR)

Coverage: Texas coast

#### NSF Award Abstract:

Hurricane Harvey is the strongest hurricane to hit the Texas coast in decades and the resulting tidal surges, flooding and terrestrial runoff have had a severe impact on the coastal ocean. The effects on the phytoplankton, the first link in the food chain, may be unprecedented. To determine how the phytoplankton community will respond to such drastic changes in salinity, nutrient inputs, and potential toxins, immediate and continuous sampling is the only way to fully capture the effects and to identify when conditions return to "normal". An automated, continuous phytoplankton imaging instrument that is deployed on the Texas coast records images of the phytoplankton and permits calculation of the abundance of different species. Together with molecular information on the genes that have been "turned on", or expressed, outcomes of this project will help determine the responses of individual types of phytoplankton. Extreme storms are expected to increase in frequency with future climate change, so the responses identified now will be valuable in predicting how such events will affect these primary producers, which in turn support most of the food webs in marine ecosystems, in the future.

High temporal resolution observations from the Imaging FlowCytobot (IFCB) have revealed that hurricanes in the Gulf of Mexico cause drastic changes in the phytoplankton community structure. The objectives of this RAPID project are: 1) to characterize the dynamics of the phytoplankton species in relation to the environmental variables along the Texas coast; 2) to assess the short and long-term changes in the phytoplankton community; and 3) to identify the strategies of the phytoplankton community for resource acquisition. To accomplish these objectives, this project will utilize IFCB time series to follow phytoplankton community structure during the recovery period from Hurricane Harvey. In addition, two RAPID response cruises (in late September and early October) to sample at 5 sites along a transect from Galveston to Port Aransas, TX. At each station, CTD profiles and water samples from surface and the chlorophyll maximum will be collected for nutrients, carbonate chemistry, and RNA sequencing for metatranscriptomic analysis. Metatranscriptomics can provide an indication of the metabolic strategies employed and functional relationships within the plankton community in response to changes in the environment. The advantage of a metatranscriptomic approach is that the entire molecular response to the environment is captured. So, while the response of phytoplankton to increased nutrient inputs from floodwater runoff is targeted, the responses to other environmental stresses (toxics, hypoxia, acidification) are also captured. Analyses of this time series using multivariate statistical techniques, such as principal component analysis (PCA), and network analysis, a powerful technique for identifying potential interactions among taxa, will provide insights on the environmental factors and metabolic responses structuring the community during the aftermath of the hurricane.

Related data from the Texas Observatory for Algal Succession Time-Series (TOAST) can be found at the following: <a href="https://toast.tamu.edu/timeline?dataset=HRR\_Cruise">https://toast.tamu.edu/timeline?dataset=HRR\_Cruise</a>

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# **Funding**

| Funding Source                           | Award       |
|--|-------------|
| NSF Division of Ocean Sciences (NSF OCE) | OCE-1760620 |

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