

# Processed CTD data (unbinned) and station information for the PE21-18 Microbial DO respir cruise (15 total casts) aboard the R/V Pelican during April and May 2021

**Website:** <https://www.bco-dmo.org/dataset/859612>

**Data Type:** Cruise Results

**Version:** 1

**Version Date:** 2021-11-08

## Project

» [CAREER: Investigating aerobic microbial respiration dynamics in coastal hypoxia](#) (Coastal O2 Respir)

Contributors	Affiliation	Role
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## Abstract

Processed CTD data (unbinned) and station information for the PE21-18 Microbial DO respir cruise (15 total casts) aboard the R/V Pelican during April and May 2021.

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## Coverage

**Spatial Extent:** N:28.8701 E:-90.4883 S:28.6074 W:-90.62202

**Temporal Extent:** 2021-04-27 - 2021-05-02

## Methods & Sampling

Fifteen CTD casts were taken during the PE21-18 cruise at 5 stations. The planned stations had the following location and depth:

Station: Lat, Long, Depth (m)  
C6C: 28.8686, -90.4903, 19.2  
D2: 28.8417, -90.8333, 15.6  
D4: 28.6083, -90.8333, 19.1  
E3: 28.6583, -91.25, 22  
F4: 28.7833, -91.6167, 24.2

The data was collected using a SeaBird 911 plus system with 12-12 liter Niskin bottles. The instruments included in the 911 plus suite are dual Temperature (SBE 3), dual Conductivity (SBE 4), and dual Oxygen (SBE 43) sensors. Additionally, the suite included an SBE 27 pH/Oxidation, Chelsea Aquatracka 3 Chl<sub>a</sub>, and a

Wetlabs CDOM. Serial numbers and calibration information is stored in the Header Information included in the .cnv files.

## Data Processing Description

The data was processed using Seabird's SBEDDataProcessing\_Win32 software. The data was converted from hexadecimal to engineering units first.

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## Data Files

File
<b>pe2118_ctd_unbinned.csv</b> (Comma Separated Values (.csv), 886.15 KB) MD5:7dd8f09bb1bdc83ff8e2e1ba959ad691 Primary data file for dataset ID 859612

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## Supplemental Files

File
<b>PE21-18 cruise plan</b> filename: Cruise_Plan_Apr2021.pdf (Portable Document Format (.pdf), 1.25 MB) MD5:3e2aa0496a8b74aaf96af059ee825ce5 Cruise plan for PE21-18 cruise with cruise tracks, dates, science operation descriptions and schematics.
<b>PE21-18 navigation plot</b> filename: nav_plot_all_topo.png (Portable Network Graphics (.png), 12.98 KB) MD5:27c079463466eb83b67cd631892e561a PE21-18 navigation plot
<b>PE21-18_unbinned_CTD</b> filename: PE21-18_unbinned_CTD.zip (ZIP Archive (ZIP), 223.56 KB) MD5:f0c89794e3438d46ced4d73c48ae507c Original Seabird CTD files (.cnv) for the dataset 859612, version 1. Sensor serial numbers and calibration information is store in the header information section.

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## Related Datasets

### IsSourceOf

Thrash, J. C. (2021) **Processed CTD data (binned) and station information for the PE21-18 Microbial DO respir cruise (15 total casts) aboard the R/V Pelican during April and May 2021.** Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2021-11-08  
doi:10.26008/1912/bco-dmo.859606.1 [[view at BCO-DMO](#)]  
*Relationship Description: The binned ctd dataset (859606) is derived from the unbinned ctd dataset (859612). The binned dataset was averaged in bins of 0.5 meters.*

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## Parameters

Parameter	Description	Units
cruise	Cruise identifier	unitless
station	Station identifier	unitless
cast_number	CTD Cast number	unitless
latitude	Latitude, south is negative	decimal degrees
longitude	Longitude, west is negative	decimal degrees
ISO_DateTime_UTC	Cast date and time (UTC) in ISO 8601 format yyyy-mm-ddTHH:MM:SSZ	unitless
depSM	Depth	m
t090C	Temperature	degrees Celsius (°C)
t190C	Temperature	degrees Celsius (°C)
c0mS_cm	Conductivity	mS/cm
c1mS_cm	Conductivity	mS/cm
sal00	Salinity	PSU
sal11	Salinity	PSU
sbeox0Mg_L	Oxygen	mg/l
sbeox1Mg_L	Oxygent	mg/l
fIC	Fluoresence	ug/l
wetCDOM	Fluoresence	ug/l
ph	pH	unitless
orp	Oxydation reduction potential	mV
flag	flag	unitless
file	file	unitless

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## Instruments

<b>Dataset-specific Instrument Name</b>	SeaBird 911 plus system
<b>Generic Instrument Name</b>	CTD Sea-Bird SBE 911plus
<b>Dataset-specific Description</b>	The data was collected using a SeaBird 911 plus system with 12-12 liter Niskin bottles. The instruments included in the 911 plus suite are dual Temperature (SBE 3), dual Conductivity (SBE 4), and dual Oxygen (SBE 43) sensors. Additionally, the suite included an SBE 27 pH/Oxidation, Chelsea Aquatracka 3 Chl_a, and a Wetlabs CDOM.
<b>Generic Instrument Description</b>	The Sea-Bird SBE 911 plus is a type of CTD instrument package for continuous measurement of conductivity, temperature and pressure. The SBE 911 plus includes the SBE 9plus Underwater Unit and the SBE 11plus Deck Unit (for real-time readout using conductive wire) for deployment from a vessel. The combination of the SBE 9 plus and SBE 11 plus is called a SBE 911 plus. The SBE 9 plus uses Sea-Bird's standard modular temperature and conductivity sensors (SBE 3 plus and SBE 4). The SBE 9 plus CTD can be configured with up to eight auxiliary sensors to measure other parameters including dissolved oxygen, pH, turbidity, fluorescence, light (PAR), light transmission, etc.). more information from Sea-Bird Electronics

<b>Dataset-specific Instrument Name</b>	Wetlabs CDOM
<b>Generic Instrument Name</b>	Fluorometer
<b>Generic Instrument Description</b>	A fluorometer or fluorimeter is a device used to measure parameters of fluorescence: its intensity and wavelength distribution of emission spectrum after excitation by a certain spectrum of light. The instrument is designed to measure the amount of stimulated electromagnetic radiation produced by pulses of electromagnetic radiation emitted into a water sample or in situ.

<b>Dataset-specific Instrument Name</b>	12-12 liter Niskin bottles
<b>Generic Instrument Name</b>	Niskin bottle
<b>Dataset-specific Description</b>	The data was collected using a SeaBird 911 plus system with 12-12 liter Niskin bottles. The instruments included in the 911 plus suite are dual Temperature (SBE 3), dual Conductivity (SBE 4), and dual Oxygen (SBE 43) sensors. Additionally, the suite included an SBE 27 pH/Oxidation, Chelsea Aquatracka 3 Chl_a, and a Wetlabs CDOM. Serial numbers and calibration information is stored in the Header Information included in the .cnv files.
<b>Generic Instrument Description</b>	A Niskin bottle (a next generation water sampler based on the Nansen bottle) is a cylindrical, non-metallic water collection device with stoppers at both ends. The bottles can be attached individually on a hydrowire or deployed in 12, 24, or 36 bottle Rosette systems mounted on a frame and combined with a CTD. Niskin bottles are used to collect discrete water samples for a range of measurements including pigments, nutrients, plankton, etc.

<b>Dataset-specific Instrument Name</b>	SBE 27 pH/Oxidation
<b>Generic Instrument Name</b>	Sea-Bird SBE 27 pH/O.R.P. sensor
<b>Generic Instrument Description</b>	The SBE 27 pH and O.R.P. (Redox) sensor combines a pressure-balanced, glass-electrode, Ag/AgCl reference probe and platinum O.R.P. electrode to provide in-situ measurements at depths to 1200 m. The replaceable pH probe is permanently sealed and is supplied with a soaker bottle attachment that prevents the reference electrode from drying out during storage. The SBE 27 is intended for use as an add-on auxiliary sensor for profiling CTDs (SBE 9plus; SBE 19, 19plus, and 19plus V2 SeaCAT; and SBE 25 and 25plus Sealogger).

<b>Dataset-specific Instrument Name</b>	dual Oxygen sensor
<b>Generic Instrument Name</b>	Sea-Bird SBE 43 Dissolved Oxygen Sensor
<b>Dataset-specific Description</b>	The data was collected using a SeaBird 911 plus system with 12-12 liter Niskin bottles. The instruments included in the 911 plus suite are dual Temperature (SBE 3), dual Conductivity (SBE 4), and dual Oxygen (SBE 43) sensors. Additionally, the suite included an SBE 27 pH/Oxidation, Chelsea Aquatracka 3 Chl_a, and a Wetlabs CDOM. Serial numbers and calibration information is stored in the Header Information included in the .cnv files.
<b>Generic Instrument Description</b>	The Sea-Bird SBE 43 dissolved oxygen sensor is a redesign of the Clark polarographic membrane type of dissolved oxygen sensors. more information from Sea-Bird Electronics

<b>Dataset-specific Instrument Name</b>	dual Temperature (SBE 3) sensor
<b>Generic Instrument Name</b>	Sea-Bird SBE-3 Temperature Sensor
<b>Dataset-specific Description</b>	The data was collected using a SeaBird 911 plus system with 12-12 liter Niskin bottles. The instruments included in the 911 plus suite are dual Temperature (SBE 3), dual Conductivity (SBE 4), and dual Oxygen (SBE 43) sensors. Additionally, the suite included an SBE 27 pH/Oxidation, Chelsea Aquatracka 3 Chl_a, and a Wetlabs CDOM. Serial numbers and calibration information is stored in the Header Information included in the .cnv files.
<b>Generic Instrument Description</b>	The SBE-3 is a slow response, frequency output temperature sensor manufactured by Sea-Bird Electronics, Inc. (Bellevue, Washington, USA). It has an initial accuracy of +/- 0.001 degrees Celsius with a stability of +/- 0.002 degrees Celsius per year and measures seawater temperature in the range of -5.0 to +35 degrees Celsius. more information from Sea-Bird Electronics

<b>Dataset-specific Instrument Name</b>	dual Conductivity (SBE 4) sensor
<b>Generic Instrument Name</b>	Sea-Bird SBE-4 Conductivity Sensor
<b>Dataset-specific Description</b>	The data was collected using a SeaBird 911 plus system with 12-12 liter Niskin bottles. The instruments included in the 911 plus suite are dual Temperature (SBE 3), dual Conductivity (SBE 4), and dual Oxygen (SBE 43) sensors. Additionally, the suite included an SBE 27 pH/Oxidation, Chelsea Aquatracka 3 Chl_a, and a Wetlabs CDOM. Serial numbers and calibration information is stored in the Header Information included in the .cnv files.
<b>Generic Instrument Description</b>	The Sea-Bird SBE-4 conductivity sensor is a modular, self-contained instrument that measures conductivity from 0 to 7 Siemens/meter. The sensors (Version 2; S/N 2000 and higher) have electrically isolated power circuits and optically coupled outputs to eliminate any possibility of noise and corrosion caused by ground loops. The sensing element is a cylindrical, flow-through, borosilicate glass cell with three internal platinum electrodes. Because the outer electrodes are connected together, electric fields are confined inside the cell, making the measured resistance (and instrument calibration) independent of calibration bath size or proximity to protective cages or other objects.

<b>Dataset-specific Instrument Name</b>	Chelsea Aquatracka 3 Chl_a
<b>Generic Instrument Name</b>	Turbidity Meter
<b>Generic Instrument Description</b>	A turbidity meter measures the clarity of a water sample. A beam of light is shown through a water sample. The turbidity, or its converse clarity, is read on a numerical scale. Turbidity determined by this technique is referred to as the nephelometric method from the root meaning "cloudiness". This word is used to form the name of the unit of turbidity, the NTU (Nephelometric Turbidity Unit). The meter reading cannot be used to compare the turbidity of different water samples unless the instrument is calibrated. Description from: <a href="http://www.gvsu.edu/wri/education/instructor-s-manual-turbidity-10.htm">http://www.gvsu.edu/wri/education/instructor-s-manual-turbidity-10.htm</a> (One example is the Orion AQ4500 Turbidimeter)

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## Deployments

### PE21-18

<b>Website</b>	<a href="https://www.bco-dmo.org/deployment/859609">https://www.bco-dmo.org/deployment/859609</a>
<b>Platform</b>	R/V Pelican
<b>Report</b>	<a href="https://datadocs.bco-dmo.org/docs/305/Coastal_O2_Respir/data_docs/Cruise_Plan_Apr2021.pdf">https://datadocs.bco-dmo.org/docs/305/Coastal_O2_Respir/data_docs/Cruise_Plan_Apr2021.pdf</a>
<b>Start Date</b>	2021-04-27
<b>End Date</b>	2021-05-02
<b>Description</b>	See more cruise information at R2R: <a href="https://www.rvdata.us/search/cruise/PE21-18">https://www.rvdata.us/search/cruise/PE21-18</a>

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## Project Information

### **CAREER: Investigating aerobic microbial respiration dynamics in coastal hypoxia (Coastal O2 Respir)**

**Coverage:** Northern Gulf of Mexico, Southern California Bight

#### *NSF Award Abstract:*

Decreasing marine dissolved oxygen (DO) is a widespread and growing global problem. Among the chief causes for coastal losses of DO are anthropogenic nutrient inputs that lead to seasonal hypoxia- DO concentrations below 2 mg/L. Microorganisms are the primary agents of oxygen removal, and although we have a basic mechanistic understanding of how nutrient enrichment combines with stratification to stimulate microbial metabolism and oxygen drawdown, we still do not know which microorganisms are ultimately responsible for oxygen consumption leading to, and during, coastal hypoxia in any particular region. Nor do we know the extent to which each hypoxic system arises from universal microbial mechanisms or whether there may be unique microorganisms and metabolic pathways involved in each locale. Hypoxic regions are increasing in number and size around the globe, amplifying the need to better understand the microbial processes responsible for oxygen consumption. In response, the project pursues an integrated research and education effort to study the microbial mechanisms of oxygen respiration in two coastal zones where DO depletion occurs: the northern Gulf of Mexico "dead zone", and the Southern California Bight. This work identifies the microorganism actively consuming oxygen in these systems, what fuels them, and whether/how those taxa respond to environmental changes. The investigators are generating microbial DO consumption rate, genetic, and taxonomic data critical to better constraining respiration models focused on water column DO depletion. In doing so, hundreds of undergraduate and students and dozens of high school STEM teachers contribute valuable data by participating in modern marine microbiological research. The project also improves integration of authentic research experiences into college and high-school classrooms in the context of a problem of global relevance.

Regions of low DO take multiple forms, from vast open ocean oxygen minimum zones (OMZs) which hover in the upper water column, to shallower coastal zones of bottom water hypoxia fueled by close proximity to fluvial inputs of human supplied nutrients. Microbial respiration is primarily responsible for the ultimate consumption of DO, and therefore understanding the microorganisms that inhabit these systems and their metabolic capabilities is critical for improving our ability to predict the timing, extent, and severity of DO depletion, and how these factors relate to environmental change. While there has been substantial research into the microbiology of OMZs, we know comparatively little about the microbial dynamics, and in particular, the microorganisms responsible for oxygen consumption, in coastal hypoxia. This integrated research and education effort is framed by the following objectives:

1. Determine the microorganisms and metabolic processes responsible for actively respiring water column DO prior to, and during, hypoxia.
2. Quantify microbial respiration rates for communities and representative water column species in both the planktonic and particle-associated fractions.
3. Integrate authentic microbiology research on a globally relevant topic into undergraduate and high school classrooms.

The work uses a combination of advanced cultivation-independent and pure culture measurements to discriminate between oxygen consumption by planktonic and particle-associated microbial fractions, as well as by communities at the surface and those in bottom waters, in two different systems of coastal DO depletion (the northern Gulf of Mexico hypoxic zone and the Southern California coastal shelf). Direct assessment of actively respiring taxa are connected with their metabolic potential, gene expression, and respiration rates. This project is determining, for the first time, the taxa actively consuming oxygen in these systems. The results also provide size fractionated respiration rates and bacterial growth efficiency (BGE) with depth across multiple sites and seasons, and importantly, also yield cell-specific respiration rates and BGE for active taxa within these systems. This data will constrain variable respiration across differing environmental conditions. The investigators are experimentally testing how alterations in environmental variables affect these respiration rates and BGEs, leading to greater predictive insight for the range of effects climate forcing will have on DO consumption. This information also facilitates comparisons between multiple marine systems to identify whether common or distinct organisms and metabolic processes are operating to remove DO.

This award reflects NSF's statutory mission and has been deemed worthy of support through evaluation using the Foundation's intellectual merit and broader impacts review criteria.

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## Funding

Funding Source	Award
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-1945279</a>

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