

# Seawater concentration data from an ocean acidification exposure experiment on adult Eastern oysters from Plum Island Sound in 2017

**Website:** <https://www.bco-dmo.org/dataset/888887>

**Data Type:** Other Field Results

**Version:** 1

**Version Date:** 2023-02-02

## Project

» [Collaborative Research: Does ocean acidification induce a methylation response that affects the fitness of the next generation in oysters?](#) (Epigenetics to Ocean)

| Contributors                     | Affiliation   | Role                               |
|----------------------------------|---|------------------------------------|
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## Abstract

Trace, minor, and major element data from adult Eastern oyster ocean acidification exposure experiments were conducted at the Ries Lab at the Northeastern University Marine Science Center on samples from Plum Island Sound in 2017. This dataset represents the phenotypic and molecular responses in the extrapallial fluid in the adult eastern oyster (*Crassostrea virginica*) exposed to experimental ocean acidification (OA) over 80 days.

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## Coverage

**Spatial Extent:** Lat:42.751636 Lon:-70.837023

**Temporal Extent:** 2017-04 - 2017-04

## Methods & Sampling

In this study, we examined the phenotypic and molecular responses in the extrapallial fluid in the adult eastern oyster (*Crassostrea virginica*) exposed to experimental ocean acidification (OA) over 80 days. The collection and culturing of *C. virginica* specimens are detailed in Downey-Wall, A.M., L.P. Cameron, B.M. Ford, E.M. McNally, Y.R. Venkataraman, S.B. Roberts, J.B. Ries, and K.E. Lotterhos. 2020. Ocean acidification induces subtle shifts in gene expression and DNA methylation in the mantle tissue of the Eastern oyster (*Crassostrea virginica*). *Frontiers in Marine Science* doi: 10.3389/fmars.2020.566419.

Extrapallial fluid (EPF) was extracted as described in Downey-Wall et al. (2020). Extrapallial fluid was extracted by inserting a sterile 5 milliliter (mL) syringe with a flexible 18-gauge polypropylene tip into the EPF cavity through the luer-lock port. The EPF was stored in 2 mL polypropylene microcentrifuge tubes with screw caps (Fisherbrand Catalog No. 02-682-558) and refrigerated at 6 degrees celsius until further analysis.

Since all tanks received water from the same header source, seawater from a subset of six tanks (2 treatment-1) was sampled for elemental analysis. Seawater samples were collected in 50-milliliter (mL) polypropylene centrifuge tubes outside of an oyster sampling timepoint near the halfway point of the experimental exposure (day 63).

### Elemental analysis

Extrapallial fluid and seawater were analyzed for trace and minor elements by inductively coupled plasma mass spectrometry (ICPMS). Liquid samples (i.e., EPF, seawater) were diluted to less than 0.05 percent total dissolved solid content with ultra-pure deionized water in 15 mL polypropylene centrifuge tubes and acidified with ultra-pure nitric acid (Fisher TraceMetal Grade Nitric Acid UN2031).

Extrapallial fluid and seawater was analyzed for a suite of 57 elements (including Ca) by ActLabs, Ontario, Canada. Liquid samples were analyzed using the ActLabs ICPMS method.

## Data Processing Description

Concentration data are data received from ActLabs:

<https://actlabs.com/geochemistry/exploration-geochemistry/4-acid-near-total-digestion/>

Concentration data were negative-corrected (i.e., for intercept correction of the calibration) by adding the lowest negative value along with a de minimis constant (0.000001) to each sample for each element that exhibited negative concentration values. Extrapallial fluid and seawater samples were then dilution-corrected. Extrapallial fluid and seawater data were converted to molarity and element-to-calcium ratios were calculated. Values were identified as outliers if the E/Ca ratio deviated by more than 103 from the mean E/Ca ratio for that element. This approach to identifying outliers removes those caused by sampling or equipment error but is more conservative than Tukey's method to identify and remove outliers.

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## Data Files

| File   |
|--|
| <b>seawater_concentration.csv</b><br>(Comma Separated Values (.csv), 64.96 KB)<br>MD5:aec1bf95bbcb50082c51904b4b2a5209<br>Primary data file for dataset 888887, version 1. |

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## Related Publications

Downey-Wall, A. M., Cameron, L. P., Ford, B. M., McNally, E. M., Venkataraman, Y. R., Roberts, S. B., Ries, J. B., & Lotterhos, K. E. (2020). Ocean Acidification Induces Subtle Shifts in Gene Expression and DNA Methylation in Mantle Tissue of the Eastern Oyster (*Crassostrea virginica*). *Frontiers in Marine Science*, 7.

<https://doi.org/10.3389/fmars.2020.566419>

*IsRelatedTo*

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## Related Datasets

### IsRelatedTo

Downey-Wall, A., Lotterhos, K., Ries, J. B., Cameron, L. (2023) **Phenotypic responses of Eastern oyster in response to variable length OA exposure conducted in summer 2017 with oysters sampled in Plum Island.** Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2023-01-20 <http://lod.bco-dmo.org/id/dataset/887553> [[view at BCO-DMO](#)]

Lotterhos, K., Ries, J. B. (2023) **Molar Ratios from an adult Eastern oyster ocean acidification exposure experiment at the Northeastern University Marine Science Center in 2017.** Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2023-02-02 doi:10.26008/1912/bco-dmo.888911.1 [[view at BCO-DMO](#)]

Lotterhos, K., Ries, J. B. (2023) **Shell Concentrations from an adult Eastern oyster ocean acidification exposure experiment on adult Eastern oysters from Plum Island Sound in 2017.** Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2023-02-02 doi:10.26008/1912/bco-dmo.888902.1 [[view at BCO-DMO](#)]

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## Parameters

| Parameter     | Description   | Units                |
|---------------|---|----------------------|
| RunID         | Unique identifier for the analysis run                            | unitless             |
| SampleID      | Unique identifier for the sample                                  | unitless             |
| Species       | type of liquid sample either seawater or extrapallial fluid (EPF) | unitless             |
| SampleType    | Species that extrapallial fluid was sampled from                  | unitless             |
| BivalveID     | Bivalve identifier  | unitless             |
| SampleWt      | weight of the sample  | grams                |
| DIWt          | weight of the deionized water added to dilute the sample          | grams                |
| NitricWeight  | weight of the nitric acid added to acidify the sample             | grams                |
| ActLabsNitric | weight of nitric acid added by ActLabs                            | grams                |
| Ag_ugL        | measured concentration of silver                                  | micrograms per liter |
| Al_ugL        | measured concentration of aluminum                                | micrograms per liter |
| As_ugL        | measured concentration of arsenic                                 | micrograms per liter |
| Ba_ugL        | measured concentration of barium                                  | micrograms per liter |
| Be_ugL        | measured concentration of beryllium                               | micrograms per liter |
| Bi_ugL        | measured concentration of bismuth                                 | micrograms per liter |
| Ca_ugL        | measured concentration of calcium                                 | micrograms per liter |
| Cd_ugL        | measured concentration of cadmium                                 | micrograms per liter |
| Ce_ugL        | measured concentration of Cerium                                  | micrograms per liter |
| Co_ugL        | measured concentration of cobalt                                  | micrograms per liter |
| Cr_ugL        | measured concentration of chromium                                | micrograms per liter |
| Cs_ugL        | measured concentration of cesium                                  | micrograms per liter |
| Cu_ugL        | measured concentration of copper                                  | micrograms per liter |
| Dy_ugL        | measured concentration of dysprosium                              | micrograms per liter |
| Er_ugL        | measured concentration of erbium                                  | micrograms per liter |

|        |  |                      |
|--------|--|----------------------|
| Eu_ugL | measured concentration of europium     | micrograms per liter |
| Fe_ugL | measured concentration of iron         | micrograms per liter |
| Ga_ugL | measured concentration of gallium      | micrograms per liter |
| Gd_ugL | measured concentration of gadolinium   | micrograms per liter |
| Ge_ugL | measured concentration of germanium    | micrograms per liter |
| Hf_ugL | measured concentration of hafnium      | micrograms per liter |
| Hg_ugL | measured concentration of mercury      | micrograms per liter |
| Ho_ugL | measured concentration of holmium      | micrograms per liter |
| In_ugL | measured concentration of indium       | micrograms per liter |
| K_ugL  | measured concentration of potassium    | micrograms per liter |
| La_ugL | measured concentration of lanthanum    | micrograms per liter |
| Li_ugL | measured concentration of lithium      | micrograms per liter |
| Lu_ugL | measured concentration of lutetium     | micrograms per liter |
| Mg_ugL | measured concentration of magnesium    | micrograms per liter |
| Mn_ugL | measured concentration of manganese    | micrograms per liter |
| Mo_ugL | measured concentration of molybdenum   | micrograms per liter |
| Na_ugL | measured concentration of sodium       | micrograms per liter |
| Nb_ugL | measured concentration of niobium      | micrograms per liter |
| Nd_ugL | measured concentration of neodymium    | micrograms per liter |
| Ni_ugL | measured concentration of nickel       | micrograms per liter |
| Pb_ugL | measured concentration of lead         | micrograms per liter |
| Pr_ugL | measured concentration of praseodymium | micrograms per liter |
| Rb_ugL | measured concentration of rubidium     | micrograms per liter |
| Sb_ugL | measured concentration of antimony     | micrograms per liter |
| Sc_ugL | measured concentration of scandium     | micrograms per liter |
| Se_ugL | measured concentration of selenium     | micrograms per liter |
| Si_ugL | measured concentration of silicon      | micrograms per liter |
| Sm_ugL | measured concentration of samarium     | micrograms per liter |
| Sn_ugL | measured concentration of tin          | micrograms per liter |
| Sr_ugL | measured concentration of strontium    | micrograms per liter |
| Ta_ugL | measured concentration of tantalum     | micrograms per liter |
| Tb_ugL | measured concentration of terbium      | micrograms per liter |
| Te_ugL | measured concentration of tellurium    | micrograms per liter |
| Th_ugL | measured concentration of thorium      | micrograms per liter |
| Ti_ugL | measured concentration of titanium     | micrograms per liter |
| Tl_ugL | measured concentration of thallium     | micrograms per liter |
| Tm_ugL | measured concentration of thulium      | micrograms per liter |
| U_ugL  | measured concentration of uranium      | micrograms per liter |
| V_ugL  | measured concentration of vanadium     | micrograms per liter |
| W_ugL  | measured concentration of tungsten     | micrograms per liter |
| Y_ugL  | measured concentration of yttrium      | micrograms per liter |

|        |                                     |                      |
|--------|-------------------------------------|----------------------|
| Yb_ugL | measured concentration of ytterbium | micrograms per liter |
| Zn_ugL | measured concentration of zinc      | micrograms per liter |
| Zr_ugL | measured concentration of zirconium | micrograms per liter |

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## Instruments

|   |   |
|---|---|
| <b>Dataset-specific Instrument Name</b> | Shiyang-III dental drill  |
| <b>Generic Instrument Name</b>          | Drill   |
| <b>Generic Instrument Description</b>   | A drill is a tool used for making round holes or driving fasteners. There are many types of drills: some are powered manually, and others use electricity (electric drill) or compressed air as the motive power. Drills with a percussive action (hammer drills) are mostly used in hard materials such as masonry (brick, concrete, and stone) or rock. Some types of hand-held drills are also used to drive screws and other fasteners. |

|   |  |
|---|--|
| <b>Dataset-specific Instrument Name</b> |  |
| <b>Generic Instrument Name</b>          | Inductively Coupled Plasma Mass Spectrometer   |
| <b>Generic Instrument Description</b>   | An ICP Mass Spec is an instrument that passes nebulized samples into an inductively-coupled gas plasma (8-10000 K) where they are atomized and ionized. Ions of specific mass-to-charge ratios are quantified in a quadrupole mass spectrometer. |

|   |   |
|---|---|
| <b>Dataset-specific Instrument Name</b> | Mettler Toledo scale                          |
| <b>Generic Instrument Name</b>          | scale   |
| <b>Dataset-specific Description</b>     | Mettler Toledo scale (precision = 0.001g)     |
| <b>Generic Instrument Description</b>   | An instrument used to measure weight or mass. |

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## Project Information

**Collaborative Research: Does ocean acidification induce a methylation response that affects the fitness of the next generation in oysters? (Epigenetics to Ocean)**

**Coverage:** Coastal Massachusetts near Nahant: 42°25'06"N 70°54'14"W

*NSF Award Abstract:*

Marine ecosystems worldwide are threatened by ocean acidification, a process caused by the unprecedented rate at which carbon dioxide is increasing in the atmosphere. Since ocean change is predicted to be rapid, extreme, and widespread, marine species may face an "adapt-or-die" scenario. However, modifications to the DNA sequence may be induced in response to a stress like ocean acidification and then inherited. Such

"epigenetic" modifications may hold the key to population viability under global climate change, but they have been understudied. The aim of this research is to characterize the role of DNA methylation, a heritable epigenetic system, in the response of Eastern oysters (*Crassostrea virginica*) to ocean acidification. The intellectual merit lies in the integrative approach, which will characterize the role of DNA methylation in the intergenerational response of oysters to ocean acidification. These interdisciplinary data, spanning from molecular to organismal levels, will provide insight into mechanisms that underlie the capacity of marine invertebrates to respond to ocean acidification and lay the foundation for future transgenerational studies. Ocean acidification currently threatens marine species worldwide and has already caused significant losses in aquaculture, especially in *Crassostrea* species. This research has broader impacts for breeding, aquaculture, and the economy. Under the investigators' "Epigenetics to Ocean" (E2O) training program, the investigators will build STEM talent in bioinformatics and biogeochemistry, expose girls in low-income school districts to careers in genomics, and advance the field through open science and reproducibility.

This research will specifically test if intermittent exposure to low pH induces a methylation response with downstream beneficial effects for biomineralization. These methylation states could be inherited and confer a fitness advantage to larvae that possess them. Phase 1 of the project will use an exposure experiment to determine the degree to which DNA methylation is altered and regulates the response to OA. Data from this experiment will be used to test the hypotheses that (i) DNA methylation, induced in the tissue of shell formation (i.e., mantle tissue), is correlated with changes in transcription and regulation of pallial fluid pH (calcifying fluid pH, measured by microelectrode), and (ii) that methylation changes induced in the mantle tissue are also induced in the germline --indicating that such changes are potentially heritable. Phase 2 of the project will use a pair-mated cross experiment to test the hypothesis that parental exposure to OA alters larval traits (calcification rate, shell structure, and polymorph mineralogy). Larvae will be generated from parents exposed to OA or control seawater, and then raised under control or OA conditions. Results will be used to (i) characterize inheritance of induced methylation states, (ii) estimate the variance in larval traits explained by genotype, non-genetic maternal/paternal effects, adult OA exposure, larval OA exposure, and parental methylome, and (iii) test the hypothesis that adult exposure alters the heritability (a quantity that predicts evolutionary response) of larval traits. Since the effects of epigenetic phenomena on estimates of heritability are highly debated, the results would advance understanding of this important issue. Because the investigators could discover that DNA methylation is a mechanism for heritable plastic responses to OA, knowledge of this mechanism would significantly improve and potentially transform predictive models for how organisms respond to global change.

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## Funding

| Funding Source   | Award                       |
|--|-----------------------------|
| <a href="#">NSF Division of Ocean Sciences (NSF OCE)</a> | <a href="#">OCE-1635423</a> |

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