

# Oyster survival difference experiments in low quality reefs in Mobile Bay, AL in September 2019

**Website:** <https://www.bco-dmo.org/dataset/892475>

**Data Type:** Other Field Results

**Version:** 1

**Version Date:** 2023-03-23

## Project

» [Collaborative Research: Keystone chemicals: Identifying general and universal molecules of fear](#) (Identifying molecules of fear)

Contributors	Affiliation	Role
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## Abstract

This dataset contains survivorship of oysters planted in low quality reefs to determine how induced defenses and habitat structural complexity influence basal prey survival. Oysters (a basal prey) induced to grow stronger shells were planted with control oysters along transects spanning the center of the reefs to outside the reefs. Oysters were left in the field for two days before individual oyster survival was assessed. Predators often produce nonconsumptive effects (NCEs) in their prey in the form of behavioral or morphological changes. Such changes often have larger or equal consequences for population dynamics as the predator directly consumes individual prey. However, it is not well understood how predators feeding across multiple trophic levels cause cascading NCEs that interact across prey trophic levels or how the prey survival benefits from these interactions change across contexts. These data help demonstrate how NCEs can influence population dynamics across space and quantify the strength of these context-dependent interactions. Data were collected by Drs. Benjamin Belgrad, Lee Smee, and Marc Weissburg from the Dauphin Island Sea Lab and Georgia Institute of Technology.

## Table of Contents

- [Coverage](#)
- [Dataset Description](#)
  - [Methods & Sampling](#)
  - [Data Processing Description](#)
- [Data Files](#)
- [Related Publications](#)
- [Related Datasets](#)
- [Parameters](#)
- [Project Information](#)
- [Funding](#)

## Coverage

**Spatial Extent:** N:30.278591 E:-88.120184 S:30.27679 W:-88.120944

**Temporal Extent:** 2019-09 - 2019-09

## Methods & Sampling

### Oyster culturing

Oysters (*Crassostrea virginica*) were cultured as spat-on-shell at the Auburn University Shellfish Laboratory (AUSL) on Dauphin Island, AL starting in late May 2019 using standard techniques (Congrove et al. 2009). Oyster larvae were settled onto sun-bleached oyster shells to create spat-on-shell. After 3 days, when oyster spat were approximately 1.0 millimeters they were exposed to either exudate from predatory blue crabs or empty cage controls in four flow-through holding tanks (length = 2.4 meters, width = 0.9 meters, water depth = 0.4 meters) supplied with unfiltered seawater pumped directly from the Gulf of Mexico. The number of spat per shell varied from ~5 - 40 and we elected to not alter initial density to mimic natural settlement during the induction period. Oysters were suspended above the tank bottom in oyster aquaculture baskets (64 x 23 x 14 centimeters) with 140 spat-covered shells per basket to prevent sediment buildup from suffocating oysters. Seven oyster baskets were present in each tank (28 total).

Spat were exposed to blue crab predator cues by holding four live caged adult blue crabs (*Callinectes sapidus*) in two of the tanks (8 crabs total), whereas the remaining two tanks contained empty cages (control) to mimic conditions where oysters regularly experience predator cues or are limited in their exposure from cues. Water volumes and crab densities were informed from established procedures (Belgrad et al. 2021). Crabs in each tank were held in four separate cages (32 centimeters x 23 centimeters x 14 centimeters) to prevent crabs from consuming the experimental oysters or each other. Every crab was fed one adult oyster daily (approximately 5.0 centimeters in length) to maximize predator cue intensity as experimental oysters would be exposed to exudates from predators and damaged conspecifics. This ensured that oysters were exposed to the most natural set of cues indicative of a predation event, which produces a strong response in oysters (Scherer et al. 2016). Crabs were replaced during the experiment as needed due to mortality. Experimental oyster baskets were rotated around the crab cages daily to reduce differences in oyster growth due to proximity to predator cues, and no differences among cages were found. The induction period was 2 months.

### Oyster survival experiment

We performed a field experiment on degraded, low-quality oyster reefs near Dauphin Island, AL to determine if the ecological relationships produced from induced defenses remain consistent across habitat quality and remained similar with laboratory experiments. Oyster spat taken from the culturing period described above (also used in the mesocosm experiment) was planted on three small (5 - 10 meters long x 3 meters wide) oyster reefs in early September 2019. Oyster reefs were within 50 meters of each other and composed of dead shell hash on a bed of sand with no mud crabs found among the hash (30°16'42.6" N; 88°07'14.7" W). No live oyster clusters were found within at least 500 meters of the site. Spat-covered shells were scraped to 10 individuals per shell to standardize predation risk. Forty-four pairs of induced and control shells were zip tied to poles and set in 11 transects total (3 - 5 transects per reef depending on its size). Transects contained four poles, with a 1-meter separation between each pole, planted in the following locations: the upper tidal zone of the reef, the lower tidal zone of the reef, the reef edge, and in the bare substrate with at least a 1-meter distance from the reef. Individual survival of all oysters was checked 48 hours after planting (n = 880 spat total; 110 spat per treatment).

## Data Processing Description

[ [table of contents](#) | [back to top](#) ]

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## Data Files

File
<b>oyster_survival_in_low_quality_reef-1.csv</b> (Comma Separated Values (.csv), 4.45 KB) MD5:0ecd31ce5a5101a736b6a5cadcc99771
Primary data file for dataset 892475, version 1.

[ [table of contents](#) | [back to top](#) ]

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## Related Publications

Belgrad, B. A., Combs, E. M., Walton, W. C., & Smee, D. L. (2021). Use of predator cues to bolster oyster

resilience for aquaculture and reef restoration. *Aquaculture*, 538, 736553.  
<https://doi.org/10.1016/j.aquaculture.2021.736553>

*Methods*

Congrove, M. S., Wesson, J. A., & Allen Jr, S. K. (2009). A practical manual for remote setting in Virginia.

*Methods*

Scherer, A. E., Lunt, J., Draper, A. M., & Smee, D. L. (2016). Phenotypic plasticity in oysters (*Crassostrea virginica*) mediated by chemical signals from predators and injured prey. *Invertebrate Biology*, 135(2), 97–107.

Portico. <https://doi.org/10.1111/ivb.12120>

*Methods*

[ [table of contents](#) | [back to top](#) ]

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## Related Datasets

### IsRelatedTo

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Belgrad, B. A., Smee, D. L., Weissburg, M. (2023) **Morphological Characteristics of Oysters from Predator Experiments at the Dauphin Island Sea Lab, AL from July to October 2020**. Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2023-03-30 doi:10.26008/1912/bco-dmo.892206.1 [[view at BCO-DMO](#)]

Belgrad, B. A., Smee, D. L., Weissburg, M. (2023) **Morphological Characteristics of Oysters from Predator Experiments at the Dauphin Island Sea Lab, AL, May-July 2019**. Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2023-03-29 doi:10.26008/1912/bco-dmo.892096.1 [[view at BCO-DMO](#)]

Belgrad, B. A., Smee, D. L., Weissburg, M. (2023) **Oyster survival differences in high-quality reefs from Skidaway Island, GA from July to October 2019**. Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2023-03-23 doi:10.26008/1912/bco-dmo.892464.1 [[view at BCO-DMO](#)]

Belgrad, B. A., Smee, D. L., Weissburg, M. (2023) **Oyster survival differences in mesocosm experiments at the Dauphin Island Sea Lab, AL between July and August 2019**. Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2023-03-29 doi:10.26008/1912/bco-dmo.892425.1 [[view at BCO-DMO](#)]

[ [table of contents](#) | [back to top](#) ]

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## Parameters

Parameter	Description	Units
shell_ID	shell identification number, each shell had 10 spat settled upon it	unitless
pole_ID	individual pole designation that shells with 10 spat on them were attached to. Each pole contained an induced and control shell (20 spat total)	unitless
transect_ID	individual transect designation; each transect ran perpendicular to reef edge with four poles	unitless
reef_ID	individual reef designation; three oyster reefs were chosen, two oyster reefs contained three transects each; the third reef contained five transects	unitless
water_depth	depth of water for each transect	meters (m)
reef_treatment	spat on poles were either planted within the reef (reef) or outside the reef (bare substrate)	unitless
pole_location	position of pole along transect; upper reef, lower reef, reef edge, bare substrate	unitless
induction_treatment	oyster shells with spat were raised in the hatchery and either continuously exposed to predator cues (induced) or received no predator cues (not induced)	unitless
a48	number of spat alive on the shell after 48 hours in the field (originally 10 spat per shell)	unitless
d48	number of spat dead on the shell after 48 hours in the field (originally 10 spat per shell)	unitless

[ [table of contents](#) | [back to top](#) ]

## Project Information

### Collaborative Research: Keystone chemicals: Identifying general and universal molecules of fear (Identifying molecules of fear)

**Coverage:** Wassaw Sound, GA, US and Dauphin Island, AL

NSF abstract:

Many prey species use chemicals released in predator urine to detect imminent danger and respond appropriately, but the identity of these 'molecules of fear' remains largely unknown. This proposal examines whether prey detect different estuarine predators using the same chemical or whether the identity of the chemical signals varies. Experiments focus on common and important estuarine prey, mud crabs and oysters, and their predators including fishes, crustaceans and marine snails. Bioactive molecules are being collected from predators and prey and characterized. The goal is to determine if there are predictive relationships between either the composition of prey flesh or the predator taxon and the signal molecule. Understanding the molecular nature of these cues can determine if there are general rules governing likely signal molecules. Once identified, investigators will have the ability to precisely manipulate or control these molecules in ecological or other types of studies. Oysters are critical to estuarine health, and they are important social, cultural and economic resources. Broader impacts of the project include training of undergraduate and graduate students from diverse backgrounds and working with aquaculture facilities and conservation managers to improve growth and survival of oysters. One response to predator cues involves creating stronger shells to deter predation. Determining the identity of cues used by oysters to detect predators can provide management options to produce oysters that either grow faster or are more resistant to predators. Project personnel is working with oystermen to increase yields of farmed oysters by managing chemical cues.

For marine prey, waterborne chemical cues are important sources of information regarding the threat of predation, thus, modulating non-consumptive effects of predation in many systems. Often such cues are produced when the predators consume the flesh of that prey. In nearly all cases, the specific bioactive molecules responsible for modulating these interactions are unknown, raising the question whether there is a

universal molecule of fear that prey respond to. Thus, the focus of the project is to determine the generality of fear-inducing metabolites released by predators and prey in estuarine food webs. The project combines metabolomics analysis of diet-derived urinary metabolites with bioassays to identify the bioactive molecules producing responses in two prey species from different taxonomic groups and trophic levels (oysters, mud crabs). Metabolites are sampled from three types of predators, fish, gastropods or crustaceans. This project aims to: 1) identify bioactive molecules produced by several common estuarine predators from different taxa; 2) compare cues from predators that induce defenses in prey vs. changes in prey behavior; and 3) contrast the identities and effects of predator-released cues with fear-inducing molecules from injured conspecifics. By identifying and contrasting the effects of waterborne molecules that induce prey responses from six predators and injured prey, this project is yielding insights into the mechanisms that mediate non-lethal predator effects, while addressing long-standing questions related to predator-prey interactions. In addition to the search of a universal molecule of fear, the experiments are exploring the role of complementary and distinct chemical information on the specificity of prey responses to different types of predators.

This award reflects NSF's statutory mission and has been deemed worthy of support through evaluation using the Foundation's intellectual merit and broader impacts review criteria.

[ [table of contents](#) | [back to top](#) ]

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## Funding

Funding Source	Award
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-1948423</a>
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-1948441</a>

[ [table of contents](#) | [back to top](#) ]