pH and total alkalinity measurements of treatment tub seawater during lab experiments conducted in spring 2022.

Website: https://www.bco-dmo.org/dataset/922037
Data Type: experimental
Version: 1
Version Date: 2024-03-24

Project
» Influence of environmental pH variability and thermal sensitivity on the resilience of reef-building corals to acidification stress (Coral Resilience)

Contributors	Affiliation	Role
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Abstract
Ocean acidification (OA) resulting from anthropogenic CO2 emissions is impairing the reproduction of marine organisms. While parental exposure to OA can protect offspring via carryover effects, this phenomenon is poorly understood in many marine invertebrate taxa. We examined how parental exposure to acidified (pH 7.40) versus ambient (pH 7.72) seawater influenced reproduction and offspring performance across six gametogenic cycles (13 weeks) in the estuarine sea anemone Nematostella vectensis. This dataset contains data resulting from measurements of treatment tub seawater conditions, including pH and total alkalinity.

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Coverage

Location: Laboratory at the University of Pennsylvania
Temporal Extent: 2022-01-26 - 2022-05-02

Dataset Description

Data generated as part of a Nematostella ocean acidfication experiment published in Glass et al., 2023. (see Related Publications). Related Zenodo datasets provides further analysis and plotting of the BCO-DMO dataset here. (see Related Dataset).

Methods & Sampling

Nematostella vectensis (Stephenson, 1935) anemones were collected from a salt marsh in Brigantine, New Jersey in the fall of 2020. Females were identified by inducing spawning, and 14 individuals that released eggs were chosen as the genotype pool for this experiment. Each female was then horizontally bisected through the body column using a razor blade, resulting in two genotypically identical individuals that were divided between the two experimental groups (ambient and acidic).

A clonal male population, also originating from the United States Atlantic coast, was obtained from the laboratory of Dr. Katerina Ragkousi (Amherst College) in the spring of 2021. The male population size was increased via bisection, resulting in a total of 20 genetically identical males for the experiment (N=10 per treatment).

All anemones were kept in 12 parts per thousand (ppt) artificial seawater (ASW; Instant Ocean Reef Crystals reef salt, Spectrum Brands, Blacksburg, VA, USA) at pH 7.7–8.1 and 18°C. The animals were maintained in a dark incubator (Boekel Scientific, Feasterville-Trevose, PA, USA) and fed approximately every other day with Artemia nauplii. The experiment was performed approximately 1–1.5 years after animal collection.

Data Processing Description

Salinity, pH, and temperature were measured and recorded approximately daily to ensure the maintenance of experimental conditions. Seawater pH was measured and recorded using a handheld pH glass electrode (Mettler Toledo, Columbus, OH, USA), which was calibrated once a week using calibration solutions (pH 7 and 10) supplied by the probe manufacturer. Salinity and temperature were measured and recorded using a handheld meter (YSI Incorporated, Yellow Springs, OH, USA), and salinity was adjusted with deionized (DI) water as needed.

Duplicate 50 mL seawater samples were collected from each tub every three days, then immediately sterilized with a 0.22 µm syringe filter (Sigma-Aldrich, Burlington, MA, USA) and stored at 4°C in conical tubes (Corning, Corning, NY, USA) until processing for total alkalinity (TA). TA was determined via titration using a Metrohm 905 Titrando (Metrohm, Herisau, Switzerland). Parameters of the carbonate system in the seawater samples [e.g. (carbonate), (bicarbonate), aragonite saturation state] were calculated from temperature, salinity, TA, and pH using the *seacarb* package in R.

BCO-DMO Processing Description

* Adjusted parameter names to comply with database requirements

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Data Files

File	
922037_v1_seawater.csv	(Comma Separated Values (.csv), 6.48 KB) MD5:3c671ab3e2ef119c8ea924906c9ba924
Primary data file for dataset ID 922037, version 1	

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Related Publications

Glass, B. H., Schmitt, A. H., Brown, K. T., Speer, K. F., & Barott, K. L. (2023). Parental exposure to ocean acidification impacts gamete production and physiology but not offspring performance in Nematostella vectensis. Biology Open, 12(3). https://doi.org/10.1242/bio.059746

Results

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Related Datasets

IsRelatedTo

Glass, B. H., Schmitt, A. H., Speer, K. F., & Barott, K. L. (2022). *Nematostella OA* [Data set]. Zenodo. https://doi.org/10.5281/ZENODO.6941530 https://doi.org/10.5281/zenodo.6941530

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Parameters

Parameter	Description	Units
Date	Date on which seawater samples were collected from experimental tubs	unitless
Treatment	Experimental treatment into which anemones were placed (ambient or acidic seawater pH)	unitless
pH	Seawater pH	Arbitrary on pH scale
mV	Conductivity of seawater in millivolts	millivolts (mV)
Salinity	Seawater salinity	unitless
Temperature	Temperature of seawater	Degrees Celsius
TA_1	Total alkalinity of sample replicate 1	umol/kg
TA_2	Total alkalinity of sample replicate 2	umol/kg
TA_average	Average total alkalinity of sample replicates	umol/kg
TA	Conversion of TA average into mol/kg	mol/kg
Alkalinity	Conversion of TA mol/kg	mol/kg*10^-3

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Instruments

Dataset-specific Instrument Name	Metrohm 905 Titrando (Metrohm, Herisau, Switzerland)
Generic Instrument Name	Metrohm 905 Titrando potentiometric titrator
Dataset-specific Description	Metrohm 905 Titrando (Metrohm, Herisau, Switzerland) for total alkalinity determination
Generic Instrument Description	The Metrohm 905 Titrando potentiometric titrator is a modular potentiometric titrator for dynamic, monotonic, and set endpoint titrations. The device includes magnetic stirrers, rod stirrers, and a titration stand. It can be connected to various dosing units which include a buret and are attached to the reagent. Operation is carried out by means of a touch-sensitive display or with high-performance PC software. Temperature is measured by a Pt1000 or NTC. Ranges of the outputs are -13 to 20 pH, -1200 to 1200 mV, and -150 to 250 deg. C (Pt1000) or -5 ro 250 deg. C (NTC). Resolutions of the outputs are 0.001 for pH, 0.1 for mV, 0.1 deg. C for temperature. The measuring interval is of 100 ms. Works in conditions from 5 to 45 deg. C and at a maximum of 80 % relative humidity.

Dataset-specific Instrument Name	Handheld pH glass electrode (Mettler Toledo, Columbus, OH, USA)
Generic Instrument Name	pH Sensor
Dataset-specific Description	Handheld pH glass electrode (Mettler Toledo, Columbus, OH, USA), calibrated once a week using calibration solutions (pH 7 and 10) supplied by the probe manufacturer
Generic Instrument Description	An instrument that measures the hydrogen ion activity in solutions. The overall concentration of hydrogen ions is inversely related to its pH. The pH scale ranges from 0 to 14 and indicates whether acidic (more H+) or basic (less H+).

Dataset-specific Instrument Name	Handheld meter (YSI Incorporated, Yellow Springs, OH, USA
Generic Instrument Name	Water Quality Multiprobe
Dataset-specific Description	Handheld meter (YSI Incorporated, Yellow Springs, OH, USA) for salnity and temperature, and salinity was adjusted with deionized (DI) water as needed
Generic Instrument Description	An instrument which measures multiple water quality parameters based on the sensor configuration.

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Project Information

Influence of environmental pH variability and thermal sensitivity on the resilience of reef-building corals to acidification stress (Coral Resilience)

Coverage: Kaneohe Bay, Oahu, HI; Heron Island, Queensland, Australia

NSF Award Abstract:

Coral reefs are incredibly diverse ecosystems that provide food, tourism revenue, and shoreline protection for coastal communities. The ability of coral reefs to continue providing these services to society is currently threatened by climate change, which has led to increasing ocean temperatures and acidity that can lead to the death of corals, the animals that build the reef framework upon which so many species depend. This project examines how temperature and acidification stress work together to influence the future health and survival of corals. The scientists are carrying out the project in Hawaii where they have found individual corals with different sensitivities to temperature stress that are living on reefs with different environmental pH conditions. This project improves understanding of how an individual coral's history influences its response to multiple stressors and helps identify the conditions that are most likely to support resilient coral communities. The project will generate extensive biological and physicochemical data that will be made freely available. Furthermore, this project supports the education and training of undergraduate and high school students and one postdoctoral researcher in marine science and coral reef ecology. Hands-on activities for high school students are being developed into a free online educational resource.

This project compares coral responses to acidification stress in populations experiencing distinct pH dynamics (high diel variability vs. low diel variability) and with distinct thermal tolerances (historically bleaching sensitive vs. tolerant) to learn about how coral responses to these two factors differ between coral species and within populations. Experiments focus on the two dominant reef builders found at these stable and variable pH reefs: Montipora capitata and Porites compressa. Individuals of each species exhibiting different thermal sensitivities (i.e., bleached vs. pigmented) were tagged during the 2015 global coral bleaching event. This system tests the hypotheses that 1) corals living on reefs with larger diel pH fluctuations have greater resilience to acidification stress, 2) coral resilience to acidification is a plastic trait that can be promoted via acclimatization, and 3) thermally sensitive corals have reduced capacity to cope with pH stress, which is exacerbated at elevated temperatures. Coral cells isolated from colonies from each environmental and bleaching history are exposed to acute pH stress and examined for their ability to recover intracellular pH in vivo using confocal microscopy, and the expression level of proteins predicted to be involved in this recovery (e.g., proton transporters) is examined via Western blot and immunolocalization. Corals from each pH history are exposed to stable and variable seawater pH in a controlled aquarium setting to determine the level of plasticity of acidification resilience and to test for pH acclimatization in this system. Finally, corals with different levels of thermal sensitivity are exposed to thermal stress and recovery, and their ability to regulate pH is examined over time. The results of these experiments help identify reef conditions that promote coral resilience to ocean acidification against the background of increasingly common thermal stress events, while advancing mechanistic understanding of coral physiology and symbiosis.

This award reflects NSF's statutory mission and has been deemed worthy of support through evaluation using the Foundation's intellectual merit and broader impacts review criteria.

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Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-1923743

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